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METHOD OF OBTAINING COMPOSITIONS
~~-DNA ENCODING A HUMAN NEUROPEPTIDE~~
~~Y/PEPTIDE YY/PANCREATIC POLYPEPTIDE RECEPTOR (Y4)~~
~~AND USES THEREOF~~
COMPRISING Y4 SPECIFIC COMPOUNDS

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Background of the Invention

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Throughout this application, various publications are referenced in parenthesis by Author and year. Full 10 citations for these references may be found at the end of the specification immediately preceding the claims. The disclosure of these publications is hereby incorporated by reference into this application to describe more fully 15 the art to which this invention pertains.

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Neuropeptides are small peptides originating from large precursor proteins synthesized by peptidergic neurons and endocrine/paracrine cells. They hold promise for treatment of neurological, psychiatric, and endocrine 20 disorders (De Wied, 1990). Often the precursors contain multiple biologically active peptides. There is great diversity of neuropeptides in the brain caused by alternative splicing of primary gene transcripts and differential precursor processing. The neuropeptide 25 receptors serve to discriminate between ligands and to activate the appropriate signals. Thus, it is expected that the receptors for neuropeptides consist of a large number of members.

30 Neuropeptide Y (NPY), a 36-amino acid peptide, is the most abundant neuropeptide to be identified in mammalian brain. NPY is an important regulator in both the central and peripheral nervous systems (Heilig et al., 1990) and influences a diverse range of physiological parameters, 35 including effects on psychomotor activity, food intake, central endocrine secretion, and vasoactivity in the cardiovascular system. High concentrations of NPY are found in the sympathetic nerves supplying the coronary, cerebral, and renal vasculature and has contributed to

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vasoconstriction. NPY binding sites have been identified in a variety of tissues, including spleen (Lundberg et al., 1988), intestinal membranes, brain (Hinson et al., 1988), aortic smooth muscle (Mihara et al., 1989), 5 kidney, testis, and placenta (Dumont et al., 1992). In addition, binding sites have been reported in a number of rat and human cell lines (eg. Y1 in SK-N-MC, MC-IXC, CHP-212, and PC12 cells; Y2 in SK-N-Be(2), CHP-234, and SMS-MSN) (Aakerlund et al., 1990; Grundemar et al., 1993).

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NPY forms a family (called the pancreatic polypeptide family) together with pancreatic polypeptide (PP) and peptide YY (PYY) which all consist of 36 amino acids and have a common tertiary structure, the so-called PP-fold 15 (Glover et al., 1985). Specific features of this family include a polyproline helix in residues 1 through 8, a β -turn in residues 9 through 14, an α -helix in residues 15 through 30, an outward-projecting C-terminus in residues 30 through 36, and a carboxy terminal amide which appears 20 to be critical for biological activity (Schwartz et al., 1990). The C-terminal amidated residue of these peptides is essential for biological activity (Wahlestedt et al., 1986). Studies with peptide fragments of NPY have indicated that multiple NPY receptor subtypes exist 25 (Wahlestedt et al., 1986). Three major NPY receptor subtypes (Y1, Y2 and Y3) have been defined by pharmacological criteria, with a fourth "atypical" Y1 receptor that has been proposed to regulate feeding behavior. The only NPY receptor which has been cloned to 30 date is the Y1 receptor gene, from mouse (Eva et al., 1992), rat (Eva et al., 1990), and human (Larhammar et al., 1992). One of the key pharmacological features which distinguish Y1 and Y2 is the fact that the Y1 receptor (and not the Y2 receptor) responds to an analog 35 of NPY modified at residues 31 and 34 ([Leu31,Pro34]NPY), whereas the Y2 receptor (and not the Y1 receptor) has high affinity for the NPY peptide carboxyl-terminal

fragment NPY-(13-36) (Wahlstedt et al., 1986; Fuhendorff et al., 1990).

Receptor genes for the other two structurally related 5 peptides, peptide YY (PYY) and pancreatic polypeptide (PP), also have not been cloned. Peptide YY occurs mainly in endocrine cells in the lower gastrointestinal tract (Bottcher et al., 1984). Receptors for PYY were first described in the rat small intestine (Laburthe et 10 al., 1986). This receptor has been defined as PYY-preferring because it displays a 5-10 fold higher affinity for PYY than for NPY (Laburthe et al., 1986; Laburthe, 1990). Recently, a cell line, PKSV-PCT, derived from the proximal tubules of kidneys, has been 15 described to express receptors for PYY (Voisin et al., 1993). Pancreatic polypeptide is predominantly located in endocrine cells of the pancreatic islets (Alumets et al., 1978). PP inhibits pancreatic exocrine secretion and gall bladder contraction (Schwartz, 1983). 20 Interestingly, PP does not appear to be synthesized in or localized to the central nervous system (Di Maggio et al., 1985), but selective PP binding sites have been found in various brain areas, such as the area postrema and adjacent nuclei, regions permeable at the blood-brain 25 barrier (Whitcomb et al., 1990). PP receptors have a much higher affinity for PP than for NPY or PYY (Inui et al., 1990). PP has been shown to bind with high affinity to binding sites on a pheochromocytoma cell line, PC12 (Schwartz et al., 1987). The rank order of affinity for 30 the pharmacologically defined receptors of NPY and related peptides are listed in Table 1.

Using an homology screening approach to clone novel NPY receptor genes, we describe here the isolation and 35 characterization of a novel NPY/PYY/PP receptor clone which we have designated Y4. The Y4 receptor appears to have a unique pharmacological profile, relative to other

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NPY-related receptors, exhibiting highest affinity for pancreatic polypeptide itself. This receptor clone will enable us to further examine the possibility of receptor diversity and the existence of multiple subtypes within 5 this family of receptors. These could then serve as invaluable tools for drug design for several pathophysiological conditions such as memory loss, depression, anxiety, epilepsy, pain, hypertension, locomotor problems, circadian rhythm disorders, 10 eating/body weight disorders, sexual/reproductive disorders, nasal congestion, diarrhea, gastrointestinal and cardiovascular disorders.

10 20 30 40 50 60 70 80 90 100 110 120 130 140 150 160 170 180 190 200 210 220 230 240 250 260 270 280 290 300 310 320 330 340 350 360 370 380 390 400 410 420 430 440 450 460 470 480 490 500 510 520 530 540 550 560 570 580 590 600 610 620 630 640 650 660 670 680 690 700 710 720 730 740 750 760 770 780 790 800 810 820 830 840 850 860 870 880 890 900 910 920 930 940 950 960 970 980 990 1000

Summary of the Invention

This invention provides an isolated nucleic acid molecule encoding a Y4 receptor.

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This invention also provides an isolated protein which is a Y4 receptor.

This invention provides a vector comprising an isolated 10 nucleic acid molecule encoding a Y4 receptor.

This invention also provides vectors such as plasmids and baculovirus comprising a nucleic acid molecule encoding a Y4 receptor, adapted for expression in a bacterial 15 cell, a yeast cell, an insect cell or a mammalian cell which additionally comprise the regulatory elements necessary for expression of the nucleic acid in the bacterial, yeast, insect or mammalian cells operatively linked to the nucleic acid encoding the Y4 receptor as to 20 permit expression thereof.

This invention provides a mammalian cell comprising nucleic acid encoding a Y4 receptor.

25 This invention provides a method for determining whether a ligand can specifically bind to a Y4 receptor which comprises contacting cell transfected with and expressing nucleic acid encoding a Y4 receptor with the ligand under conditions permitting binding of ligands to such Y4 30 receptor, and detecting the presence of any of the ligand bound to a Y4 receptor, thereby determining whether the ligand binds specifically to a Y4 receptor.

This invention also provides a method for determining 35 whether a ligand is a Y4 receptor agonist which comprises contacting a cell transfected with and expressing nucleic acid encoding a Y4 receptor with the ligand under

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conditions permitting the activation of a Y4 receptor functional response from the cell, and detecting by means of a bioassay, such as a second messenger response, an increase in Y4 receptor activity, thereby determining 5 whether the ligand is a Y4 receptor agonist.

This invention further provides a method for determining whether a ligand is a Y4 receptor antagonist which comprises contacting a cell transfected with and 10 expressing nucleic acid encoding a Y4 receptor with the ligand under conditions permitting the activation of a functional Y4 receptor response, and detecting by means of a bioassay, such as a second messenger response, a decrease in Y4 receptor activity, and thereby determining 15 whether the ligand is a Y4 receptor antagonist.

This invention further provides a method of screening drugs to identify drugs which specifically bind to a Y4 receptor which comprises contacting a cell transfected 20 with and expressing nucleic acid encoding a Y4 receptor with a plurality of drugs, and determining those drugs which bind to the cell, thereby identifying drugs which specifically bind to a Y4 receptor.

25 This invention also provides a method of screening drugs to identify which act as agonists of a Y4 receptor which comprises contacting a cell transfected with and expressing a Y4 receptor with a plurality of drugs under conditions permitting the activation of a functional Y4 30 receptor response, and determining those drugs which activate the receptor in the cell using a bioassay such as a second messenger assay, thereby identifying drugs Y4 receptor agonists.

35 This invention also provides a method of screening drugs to identify drugs which act as antagonists of a Y4 receptor which comprises contacting a cell transfected

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with and expressing a Y4 receptor with a plurality of drugs in the presence of a known human Y4 receptor agonist, such as PP, under conditions permitting the activation of a functional Y4 receptor response and

5 determining those drugs which inhibit the activation of the receptor in the cell using a bioassay, such as a second messenger assay, thereby identifying drugs which act as antagonists of a Y4 receptor.

10 This invention provides a nucleic acid probe comprising a nucleic acid molecule of at least 15 nucleotides capable of specifically hybridizing with a unique sequence included within the sequence of a nucleic acid molecule encoding a Y4 receptor.

15 This invention also provides a method of detecting expression of the Y4 receptor on the surface of a cell by detecting the presence of mRNA coding for a Y4 receptor which comprises obtaining total mRNA from the cell and

20 contacting the mRNA so obtained with a nucleic acid probe comprising a nucleic acid molecule of at least 15 nucleotides capable of specifically hybridizing with a unique sequence included within the sequence of a nucleic acid molecule encoding a Y4 receptor under hybridizing

25 conditions, detecting the presence of mRNA hybridized to the probe, and thereby detecting the expression of the Y4 receptor by the cell.

30 This invention provides an antisense oligonucleotide having a sequence capable of hybridizing specifically an mRNA molecule which encodes a Y4 receptor so as to prevent translation of the mRNA molecule.

35 This invention provides an antibody directed to a Y4 receptor.

This invention provides a transgenic nonhuman mammal

expressing nucleic acid encoding a Y4 receptor. This invention further provides a transgenic nonhuman mammal whose genome comprises antisense DNA complementary to DNA encoding a Y4 receptor so placed as to be transcribed 5 into antisense mRNA which is complementary to mRNA encoding a Y4 receptor and which hybridizes to mRNA encoding a Y4 receptor thereby reducing its translation..

This invention provides a method of determining the 10 physiological effects of expressing varying levels of Y4 receptors which comprises producing a transgenic nonhuman animal whose levels of Y4 receptor expression are varied by use of an inducible promoter which regulates Y4 receptor expression.

15 This invention also provides a method of determining the physiological effects of expressing varying levels of Y4 receptors which comprises producing a panel of transgenic nonhuman animals each expressing a different amount of Y4 20 receptor.

This invention provides a method for diagnosing a predisposition to a disorder associated with the activity of a specific human Y4 receptor allele which comprises:
25 a. obtaining nucleic acid of subjects suffering from the disorder; b. performing a restriction digest of the nucleic acid with a panel of restriction enzymes; c. electrophoretically separating the resulting nucleic acid fragments on a sizing gel; d. contacting the resulting 30 gel with a nucleic acid probe capable of specifically hybridizing to nucleic acid encoding a Y4 receptor and labelled with a detectable marker; e. detecting labelled bands which have hybridized to the nucleic acid encoding a Y4 receptor labelled with a detectable marker to 35 create a unique band pattern specific to the DNA of subjects suffering from the disorder; f. preparing nucleic acid obtained for diagnosis by steps a-e; and g.

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comparing the unique band pattern specific to the nucleic acid of subjects suffering from the disorder from step e and the nucleic acid obtained for diagnosis from step f to determine whether the patterns are the same or 5 different and to diagnose thereby predisposition to the disorder if the patterns are the same.

This invention provides a method of preparing the purified, isolated Y4 receptor which comprises a)

10 constructing a vector adapted for expression in a cell which comprises the regulatory elements necessary for the expression of nucleic acid in the cell operatively linked to the nucleic acid encoding a Y4 receptor as to permit expression thereof, wherein the cell is selected from the

15 group consisting of bacterial cells, yeast cells, insect cells and mammalian cells; b) inserting the vector of step a in a suitable host cell; c) incubating the cells of step b under conditions allowing the expression of a Y4 receptor; d) recovering the receptor so produced; and e)

20 purifying the receptor so recovered, thereby preparing an isolated, purified Y4 receptor.

This invention also provides a method of preparing the isolated Y4 receptor which comprises inserting nucleic acid encoding Y4 receptor in a suitable vector, inserting the resulting vector in a suitable host cell, recovering the receptor produced by the resulting cell, and purifying the receptor so recovered.

30 This invention provides an antisense oligonucleotide having a sequence capable of binding specifically with any sequences of an mRNA molecule which encodes the Y4 receptor so as to prevent translation of mRNA molecules which encode the Y4 receptor.

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This invention also provides a transgenic nonhuman mammal expressing DNA encoding a human Y4 receptor.

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This invention further provides a transgenic nonhuman mammal comprising a homologous recombination knockout of the native Y4 receptor.

5 This invention also provides a method of determining the physiological effects of expressing varying levels of a receptor which comprises producing a transgenic nonhuman animal whose levels of human Y4 receptor expression are varied by use of an inducible promoter which regulates
10 receptor expression.

This invention also provides a method of determining the physiological effects of expressing varying levels of a human Y4 receptor which comprises producing a panel of
15 transgenic nonhuman animals each expressing a different amount of the receptor.

This invention further provides a transgenic nonhuman mammal whose genome comprises antisense DNA complementary
20 to DNA encoding a human Y4 receptor so placed as to be transcribed into antisense mRNA which is complementary to mRNA encoding the receptor and which hybridizes to mRNA encoding the receptor thereby preventing its translation.

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This invention provides a method for determining whether a ligand not known to be capable of binding to the Y4 receptor can bind to the receptor which comprises contacting a mammalian cell comprising an isolated DNA
30 molecule encoding the Y4 receptor with the ligand under conditions permitting binding of ligands known to bind to the receptor, detecting the presence of any of the ligand bound to the Y4 receptor, and thereby determining whether the ligand binds to the Y4 receptor.

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Brief Description of the FiguresFigure 1

5 Nucleotide Sequence and Dduced Amino Acid Sequence of a Novel Human hp25a Neuropeptide Receptor (Sequence I.D. Nos. 1 and 2). Nucleotides are presented in the 5' to 3' orientation and the coding region is numbered starting from the initiating methionine and ending in the
10 termination codon. Dduced amino acid sequence by translation of a long open reading frame is shown, along with the 5' and 3' untranslated regions. Numbers in the left and right margins represent nucleotide (top line) and amino acid (bottom line) numberings, starting with
15 the first position as the adenosine (A) and the initiating methionine (M), respectively.

Figure 2

Sequence Alignment of the Human hp25a clone with human
20 Y1, rat Y1, and mouse Y1 receptor genes. The deduced amino acid sequence of the human hp25a (Y4) receptor (first line, ^(SEQ ID NO:2) from the starting methionine (M) to the stop codon (*), is aligned with the human Y1 receptor clone (^(SEQ ID NO:34) Larhammar et al., 1992), rat Y1 receptor clone (^(SEQ ID NO:35) ^(SEQ ID NO:25) (Eva et al., 1990), and mouse Y1 receptor clone (^(SEQ ID NO:36) (Eva et al., 1992). Hyphens represent added spaces necessary for proper alignment. Gray shading indicates residues in receptor clones which are identical to hp25a. Numbers above amino acid sequences correspond to amino acid
30 positions of hp78a, starting with the initiating methionine (M) and ending with the termination codon (*), and including spaces to account for proper alignment. Solid bars above the sequence indicate the seven putative transmembrane (TM) spanning regions (TM I - VII).
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Figure 3. Nucleotide sequence and deduced amino acid sequence of the rat Y4 receptor encoded by rs16b

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(SEQ ID NOS: 27 and 28)

(Sequence 1.5. Nos and). Nucleotides are presented in the 5' to 3' orientation and the coding region is numbered starting from the putative initiating methionine and ending in the termination codon. Deduced amino acid sequence by translation of a long open reading frame is shown, along with 5' and 3' untranslated regions. The amino acid sequence is represented using single-letter abbreviations.

10 Figure 4. Alignment of rat and human Y4 receptors. Predicted amino acid sequences of the rat Y4 receptor (Y4rat) and human Y4 receptor (Y4hum) are shown; the sequences are 75% identical overall and 84% identical in the transmembrane domains. Single letter abbreviations 15 for amino acids are shown. The seven putative transmembrane (TM) spanning regions (TM I - VII) are indicated by brackets above the sequence.

Figure 5

20 Equilibrium binding of ^{125}I -PYY to membranes from COS-7 cells transiently expressing hp25a receptors. Membranes were incubated with ^{125}I -PYY for the times indicated, in the presence or absence of 100 nM human PP. Specific binding, B , was plotted against time, t , to obtain the 25 maximum number of equilibrium binding sites, B_e , and observed association rate, K_{obs} ; according to the equation, $B = B_e * (1 - e^{-(k_{obs} * t)})$. Binding is shown as the percentage of total equilibrium binding, B_e , determined by nonlinear regression analysis. Data are representative 30 of three independent experiments, with each point measured in triplicate.

Figure 6A

Saturable equilibrium binding of ^{125}I -PYY to membranes from 35 COS-7 cells transiently expressing hp25a receptors. Membranes were incubated with ^{125}I -PYY ranging in concentration from 0.003 nM to 2 nM, in the presence or

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absence of 100 nM human PP.

Figure 6B

Specific binding of the ^{125}I -PYY to membranes from COS-7
5 cells transiently expressing hp25a receptors under the
conditions described in Figure 6A was plotted against the
free ^{125}I -PYY concentration, [L], to obtain the maximum
number of saturable binding sites, B_{\max} , and the ^{125}I -PYY
equilibrium dissociation constant, K_d , according to the
10 binding isotherm, $B = B_{\max}[L]/([L] + K_d)$. Specific binding
is shown for data from a representative of four
independent experiments, with each point measured in
quadruplicate.

15 Figure 7. Competitive displacement of ^{125}I -PYY from COS-7
cells transiently expressing hp25a receptors. Membranes
were incubated with ^{125}I -PYY and increasing concentrations
of peptide competitors. IC_{50} values corresponding to 50%
displacement were determined by nonlinear regression
20 analysis and converted to K_i values according to the
equation, $K_i = IC_{50}/(1 + [L]/K_d)$, where [L] is the ^{125}I -PYY
concentration and K_d is the equilibrium dissociation
constant of ^{125}I -PYY. Data are representative of at least
two independent experiments, with each point measured
25 once or in duplicate. Rank orders of affinity for these
and other compounds are listed separately in Table 2.

Figure 8. Inhibition of forskolin-stimulated cAMP
30 accumulation in intact LM(tk-) cells stably expressing
the human Y4 receptor. Functional data were derived
from radioimmunoassay of cAMP in LM(tk-) cells stimulated
with 10 μM forskolin over a 5 minute period. Human PP was
tested for agonist activity at concentrations ranging
35 from 0.03 pM to 0.3 μM over the same period. Data were
fit to a four parameter logistic equation by nonlinear
regression. The data shown are representative of three

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independent experiments.

Figures 9A and 9B. Figure 9A. Stimulation of intracellular free calcium concentration in intact LM(tk-) cells stably expressing the human Y4 receptor. Representative time course. Functional data were derived from Fura-2/AM fluorescence in LM(tk-) cells stimulated with 100 nM human PP (open squares) or 100 nM human NPY (closed squares) at the time indicated by the arrow. The data shown are representative of two independent experiments. Figure 9B. Concentration/response curve. Data were fit to a four parameter logistic equation by nonlinear regression."

Detailed Description of the Invention

Throughout this application, the following standard
5 abbreviations are used to indicate specific nucleotide
bases:

C = cytosine A = adenine
T = thymine G = guanine

This invention provides isolated nucleic acid molecules
10 which encode Y4 receptors. In one embodiment the Y4
receptor encoded is a rat Y4 receptor. In another
embodiment, the Y4 receptor encoded is a human Y4
receptor. In an embodiment, the isolated nucleic acid
molecule encodes a Y4 receptor being characterized by an
15 amino acid sequence in the transmembrane region, wherein
the amino acid sequence has 60% homology or higher to the
amino acid sequence in the transmembrane region of the
human Y4 receptor shown in Figure 2. In another
embodiment, the Y4 receptor has substantially the same
20 amino acid sequence as the human Y4 receptor as described
in Figure 1. In yet another embodiment, the Y4 receptor
has substantially the same amino acid sequence as the rat
Y4 receptor as described in Figure 3. In another
embodiment, the Y4 receptor has the amino acid sequence
25 as shown in Figure 1. In another embodiment, the Y4
receptor has the amino acid sequence as shown in Figure
3. As used herein, the term Y4 receptor encompasses any
amino acid sequence, polypeptide or protein having
substantially the same pharmacology provided subject
30 human Y4 receptor as shown in Tables 1-3 and Table 6 and
Figures 5-7. As described herein the human Y4 receptor
has a pharmacological profile that differs from any known
neuropeptide Y receptor subtype (i.e. Y1, Y2 and Y3),
Neuropeptide YY receptor, and pancreatic polypeptide
35 receptor, and is therefore designated as the human Y4
receptor.

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The only NPY receptor which has been cloned to date is the Y1 receptor gene, from mouse (Eva et al., 1992), rat (Eva et al., 1990), and human (Larhammar et al., 1992). The Y4 receptor's greatest homology with any known 5 receptor disclosed in the Genbank/EMBL databases is a 42% overall amino acid identity with the human Y1 receptor.

This invention provides an isolated nucleic acid molecule encoding a Y4 receptor. In one embodiment, the Y4 10 receptor is a human Y4 receptor. In another embodiment, the Y4 receptor is a rat Y4 receptor. As used herein, the term "isolated nucleic acid molecule" means a nucleic acid molecule that is a molecule in a form which does not occur in nature. Examples of such an isolated nucleic 15 acid molecule are an RNA, cDNA, or isolated genomic DNA molecule encoding a Y4 receptor. One means of isolating a human Y4 receptor is to probe a human genomic library with a natural or artificially designed DNA probe, using methods well known in the art. DNA probes derived from 20 the human receptor gene Y4 are particularly useful probes for this purpose. DNA and cDNA molecules which encode human Y4 receptors may be used to obtain complementary genomic DNA, cDNA or RNA from human, mammalian or other animal sources, or to isolate related cDNA or genomic 25 clones by the screening of cDNA or genomic libraries, by methods described in more detail below. Transcriptional regulatory elements from the 5' untranslated region of the isolated clones, and other stability, processing, transcription, translation, and tissue 30 specificity-determining regions from the 3' and 5' untranslated regions of the isolated genes are thereby obtained. Examples of a nucleic acid molecule are an RNA, cDNA, or isolated genomic DNA molecule encoding a Y4 receptor. Such molecules may have coding sequences such 35 as the coding sequences shown in Figures 1 or 3. The DNA molecule of Figure 1 encodes the amino acid sequence of a human Y4 receptor protein, while the DNA molecule of

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Figure 3 encodes the amino acid sequence of the rat Y4 receptor.

This invention further provides a cDNA molecule encoding 5 a Y4 receptor having a coding sequence substantially the same as the coding sequence shown in Figures 1 and 3. This molecule is obtained by the means described above.

This invention also provides an isolated protein which is 10 a Y4 receptor. In one embodiment, the Y4 receptor is a human Y4 receptor. In another embodiment, the Y4 receptor is a rat Y4 receptor. As used herein, the term "isolated protein" means a protein molecule free of other cellular components. An example of such a protein is an 15 isolated protein having substantially the same amino acid sequence as the amino acid sequence shown in Figure 1 which is a human Y4 receptor or the amino acid sequence shown in Figure 3 which is a rat Y4 receptor. One means for obtaining isolated Y4 receptor is to express DNA 20 encoding the receptor in a suitable host, such as a bacterial, yeast, insect or mammalian cell, using methods well known in the art, and recovering the receptor protein after it has been expressed in such a host, again using methods well known in the art. The receptor may 25 also be isolated from cells which express it, in particular from cells which have been transfected with the expression vectors described below in more detail.

This invention provides vectors comprising isolated 30 nucleic acid molecules such as DNA, RNA, or cDNA encoding a Y4 receptor. In one embodiment the Y4 receptor is a human Y4 receptor. In another embodiment the Y4 receptor is a rat Y4 receptor. Examples of vectors are viruses such as bacteriophages (such as phage lambda), animal 35 viruses (such as Herpes virus, Murine Leukemia virus, and Baculovirus), cosmids, plasmids (such as pUC18, available from Pharmacia, Piscataway, NJ), and other recombination

vectors. Nucleic acid molecules are inserted into vector genomes by methods well known in the art. For example, insert and vector DNA can both be exposed to a restriction enzyme to create complementary ends on both 5 molecules which base pair with each other and are then ligated together with a ligase. Alternatively, linkers can be ligated to the insert DNA which correspond to a restriction site in the vector DNA, which is then digested with the restriction enzyme which cuts at that 10 site. Other means are also available. Specific examples of such plasmids are plasmids comprising cDNA having a coding sequence substantially the same as the coding sequence shown in Figure 1 and designated clone hp25a (Seq. I.D. No. 1) or the coding sequence shown in Figure 15 3 and designated clone rs16b (Sequence I.D. No. 27).

This invention also provides vectors comprising DNA molecules encoding Y4 receptors, adapted for expression in a bacterial cell, a yeast cell, an insect cell or a 20 mammalian cell which additionally comprise the regulatory elements necessary for expression of the DNA in the bacterial, yeast, insect or mammalian cells operatively linked to the DNA encoding a Y4 receptor as to permit expression thereof. DNA having coding sequences 25 substantially the same as the coding sequence shown in Figure 1 may usefully be inserted into the vectors to express human Y4 receptors. DNA having coding sequences substantially the same as the coding sequence shown in Figure 3 may usefully be inserted into the vectors to 30 express rat Y4 receptors. Regulatory elements required for expression include promoter sequences to bind RNA polymerase and transcription initiation sequences for ribosome binding. For example, a bacterial expression vector includes a promoter such as the lac promoter and 35 for transcription initiation the Shine-Dalgarno sequence and the start codon AUG (Maniatis, et al., Molecular Cloning, Cold Spring Harbor Laboratory, 1982).

Similarly, a eukaryotic expression vector includes a heterologous or homologous promoter for RNA polymerase II, a downstream polyadenylation signal, the start codon AUG, and a termination codon for detachment of the 5 ribosome. Furthermore, an insect expression vector, such as recombinant Baculovirus, uses the polyhedrin gene expression signals for expression of the inserted gene in insect cells. Such vectors may be obtained commercially or assembled from the sequences described by methods well 10 known in the art, for example the methods described above for constructing vectors in general. Expression vectors are useful to produce cells that express the receptor. Certain uses for such cells are described in more detail below.

15 This invention further provides a plasmid adapted for expression in a bacterial, yeast, insect, or, in particular, a mammalian cell which comprises a DNA molecule encoding a Y4 receptor and the regulatory 20 elements necessary for expression of the DNA in the bacterial, yeast, insect, or mammalian cell operatively linked to the DNA encoding a Y4 receptor as to permit expression thereof. Some plasmids adapted for expression in a mammalian cell are pSVL (available from Pharmacia, 25 Piscataway, NJ) and pcEXV-3 (Miller J. and Germain R.N., J. Exp. Med. 164:1478 (1986)). A specific example of such plasmid is a plasmid adapted for expression in a mammalian cell comprising cDNA having coding sequences substantially the same as the coding sequence shown in 30 Figure 1 and the regulatory elements necessary for expression of the DNA in the mammalian cell which is designated pcEXV-Y4 and deposited under ATCC Accession No. 75631. Another example of such plasmid is a plasmid adapted for expression in a mammalian cell comprising 35 cDNA having coding sequences substantially the same as the coding sequence shown in Figure 3 and the regulatory elements necessary for expression of the DNA in the

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a
mammalian cell which is designated pcEXV-rY4 and deposited under ATCC Accession number 75984. Those skilled in the art will readily appreciate that numerous plasmids adapted for expression in a mammalian cell which 5 comprise DNA encoding Y4 receptors and the regulatory elements necessary to express such DNA in the mammalian cell may be constructed utilizing existing plasmids and adapted as appropriate to contain the regulatory elements necessary to express the DNA in the mammalian cell. The 10 plasmids may be constructed by the methods described above for expression vectors and vectors in general, and by other methods well known in the art.

15 The deposit discussed supra, and the other deposits discussed herein, were made pursuant to, and in satisfaction of, the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, 20 Rockville, Maryland 20852.

This invention provides a cell comprising a nucleic acid encoding a Y4 receptor, such as a mammalian cell comprising a plasmid adapted for expression in a mammalian cell, which comprises a nucleic acid molecule 25 encoding a Y4 receptor, the protein encoded thereby is expressed on the cell surface, and the regulatory elements necessary for expression of the nucleic acid in the mammalian cell operatively linked to the nucleic acid 30 encoding a Y4 receptor as to permit expression thereof. Numerous mammalian cells may be used as hosts, including, for example, the mouse fibroblast cell NIH-3T3, CHO cells, HeLa cells, LM(tk-) cells, Y1 cells, etc. Expression plasmids such as that described supra may be 35 used to transfect mammalian cells by methods well known in the art such as calcium phosphate precipitation, or DNA encoding these Y4 receptors may be otherwise

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introduced into mammalian cells, e.g., by microinjection, to obtain mammalian cells which comprise DNA, e.g., cDNA or a plasmid, encoding either Y4 receptor. In one embodiment, the LM(tk-) cell is designated L-hY4-3 (ATCC 5 Accession No. CRL-1179). In another embodiment, the NIH-3t3 cell is designated N-hY4-5 (ATCC Accession No. CRL-1178)

This invention provides a method for determining whether a ligand can specifically bind to a Y4 receptor which 10 comprises contacting a cell transfected with and expressing nucleic acid encoding the Y4 receptor with the ligand under conditions permitting binding of ligands to such receptor, and detecting the presence of any such ligand bound specifically to the Y4 receptor, thereby 15 determining whether the ligand binds specifically to a Y4 receptor. In one embodiment, the Y4 receptor is a human Y4 receptor. In another embodiment, the Y4 receptor is a rat Y4 receptor.

20 This invention provides a method for determining whether a ligand can specifically bind to a Y4 receptor which comprises contacting a cell transfected with and expressing nucleic acid encoding the Y4 receptor with the ligand under conditions permitting binding of ligands to 25 such receptor, and detecting the presence of any such ligand bound specifically to the Y4 receptor, thereby determining whether the ligand binds specifically to a Y4 receptor, wherein the Y4 receptor is characterized by an amino acid sequence in the transmembrane region, wherein 30 the amino acid sequence has 60% homology or higher to the amino acid sequence in the transmembrane region of the human Y4 receptor shown in Figure 2. In one embodiment, the Y4 receptor is a human Y4 receptor. In another embodiment, the Y4 receptor is a rat Y4 receptor.

35 This invention provides a method for determining whether a ligand can bind specifically to a Y4 receptor which

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comprises preparing a cell extract from cells transfected with and expressing nucleic acid encoding a Y4 receptor, isolating a membrane fraction from the cell extract, contacting the ligand with the membrane fraction under 5 conditions permitting binding of ligands to such receptor, and detecting the presence of any ligand bound to the Y4 receptor, thereby determining whether the compound is capable of specifically binding to a Y4 receptor. In one embodiment, the Y4 receptor is a human 10 Y4 receptor. In another embodiment, the Y4 receptor is a rat Y4 receptor.

This invention provides a method for determining whether a ligand is a Y4 receptor agonist which comprises 15 contacting a cell transfected with and expressing nucleic acid encoding a Y4 receptor with the ligand under conditions permitting the activation of a functional Y4 receptor response from the cell, and detecting by means of a bioassay, such as a second messenger response, an 20 increase in Y4 receptor activity, thereby determining whether the ligand is a Y4 receptor agonist. In one embodiment, the Y4 receptor is a human Y4 receptor. In another embodiment, the Y4 receptor is a rat Y4 receptor.

25 This invention provides a method for determining whether a ligand is a Y4 receptor agonist which comprises preparing a cell extract from cells transfected with and expressing nucleic acid encoding a Y4 receptor, isolating a membrane fraction from the cell extract, contacting the 30 membrane fraction with the ligand under conditions permitting the activation of a functional Y4 receptor response, and detecting by means of a bioassay, such as a second messenger response, an increase in Y4 receptor activity, thereby determining whether the ligand is a Y4 receptor agonist. In one embodiment, the Y4 receptor is a human Y4 receptor. In another embodiment, the Y4 35 receptor is a rat Y4 receptor.

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This invention provides a method for determining whether a ligand is a Y4 receptor antagonist which comprises contacting a cell transfected with and expressing nucleic acid encoding a Y4 receptor with the ligand in the presence of a known Y4 receptor agonist, such as PP, under conditions permitting the activation of a functional Y4 receptor response and detecting by means of a bioassay, such as a second messenger response, a decrease in Y4 receptor activity, thereby determining whether the ligand is a Y4 receptor antagonist. In one embodiment, the Y4 receptor is a human Y4 receptor. In another embodiment, the Y4 receptor is a rat Y4 receptor.

This invention provides a method for determining whether a ligand is a Y4 receptor antagonist which comprises preparing a cell extract from cells transfected with and expressing nucleic acid encoding a Y4 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with the ligand in the presence of a known Y4 receptor agonist, such as PP, under conditions permitting the activation of a functional Y4 receptor response and detecting by means of a bioassay, such as a second messenger response, a decrease in Y4 receptor activity, thereby determining whether the ligand is a Y4 receptor antagonist. In one embodiment, the Y4 receptor is a human Y4 receptor. In another embodiment, the Y4 receptor is a rat Y4 receptor.

In one embodiment of the above-described methods, the ligand is not previously known.

This invention provides a Y4 receptor agonist detected by the above-described method. This invention provides a Y4 receptor antagonist detected by the above-described method.

As used herein, the term "agonist" means any ligand

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capable of increasing Y4 receptor activity. As used herein, the term "antagonist" means any ligand capable of decreasing Y4 receptor activity.

5 In one embodiment of the above-described methods, the cell is a mammalian cell. In a further embodiment, the cell is non-neuronal in origin. In another embodiment, the nonneuronal cell is a COS-7 cell, a CHO cell, an NIH-3T3 cell or an LM(tk-) cell.

10

One method for determining whether a ligand is capable of binding to the human Y4 receptor comprises contacting a transfected nonneuronal cell (i.e. a cell that does not naturally express any type of NPY, PP, or PYY receptor, 15 thus will only express such a receptor if it is transfected into the cell) expressing a Y4 receptor on its surface, or contacting a membrane preparation derived from such a transfected cell, with the ligand under conditions which are known to prevail, and thus to be 20 associated with, in vivo binding of the ligands to a Y4 receptor, detecting the presence of any of the ligand being tested bound to the Y4 receptor on the surface of the cell, and thereby determining whether the ligand binds to, activates or inhibits the activation of the Y4 25 receptor. A response system for detecting the activation or inhibition of activation of the Y4 receptor is obtained by transfection of isolated DNA into a suitable host cell containing the desired second messenger system such as phosphoinositide hydrolysis, adenylate cyclase, 30 guanylate cyclase or ion channels. Such a suitable host cell system is isolated from pre-existing cell lines, or can be generated by inserting appropriate components of second messenger systems into existing cell lines. Such a transfection system provides a complete response system 35 for investigation or assay of the activity of Y4 receptors with ligands as described above. Transfection systems are useful as living cell cultures for

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competitive binding assays between known or candidate drugs and ligands which bind to the receptor and which are labeled by radioactive, spectroscopic or other reagents. Membrane preparations containing the receptor 5 isolated from transfected cells are also useful for these competitive binding assays. Functional assays of second messenger systems or their sequelae in transfection systems act as assays for binding affinity and efficacy in the activation of receptor function. A transfection 10 system constitutes a "drug discovery system" useful for the identification of natural or synthetic compounds with potential for drug development that can be further modified or used directly as therapeutic compounds to activate or inhibit the natural functions of the Y4 15 receptor. The transfection system is also useful for determining the affinity and efficacy of known drugs at the Y4 receptor sites.

This invention also provides a method of screening drugs 20 to identify drugs which specifically bind to a Y4 receptor on the surface of a cell which comprises contacting a cell transfected with and expressing nucleic acid encoding a Y4 receptor with a plurality of drugs, and determining those drugs which bind to the cell, 25 thereby identifying drugs which specifically bind to a Y4 receptor.

This invention also provides a method of screening drugs to identify drugs which specifically bind to a Y4 30 receptor on the surface of a cell which comprises preparing a cell extract from cells transfected with and expressing nucleic acid encoding a Y4 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with a plurality of drugs, and 35 determining those drugs which bind to the membrane fraction, thereby identifying drugs which specifically bind to a Y4 receptor.

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This invention also provides a method of screening drugs to identify drugs which act as Y4 receptor agonists which comprises contacting a cell transfected with and expressing nucleic acid encoding a Y4 receptor with a 5 plurality of drugs under conditions permitting the activation of a functional Y4 receptor response, determining those drugs which activate the Y4 receptor in the cell using a bioassay, such as a second messenger assay, thereby identifying drugs which act as Y4 receptor 10 agonists.

This invention also provides a method of screening drugs to identify drugs which act as Y4 receptor agonists which comprises preparing a cell extract from cells transfected 15 with and expressing nucleic acid encoding a Y4 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with a plurality of drugs under conditions permitting the activation of a functional Y4 receptor response, determining those drugs 20 which activate the Y4 receptor in the cell using a bioassay, such as a second messenger assay, thereby identifying drugs which act as Y4 receptor agonists.

This invention also provides a method of screening drugs 25 to identify drugs which act as Y4 receptor antagonists which comprises contacting a cell transfected with and expressing DNA encoding a Y4 receptor with a plurality of drugs in the presence of a known Y4 receptor agonist, such as PP, under conditions permitting the activation of 30 a functional Y4 receptor response, and determining those drugs which inhibit the activation of the Y4 receptor in the cell using a bioassay, such as a second messenger assay, thereby identifying drugs which act as Y4 receptor antagonists.

35 This invention also provides a method of screening drugs to identify drugs which act as Y4 receptor antagonists which comprises preparing a cell extract from cells

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transfected with and expressing nucleic acid encoding a Y4 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with a plurality of drugs in the presence of a known Y4 receptor 5 agonist, such as PP, under conditions permitting the activation of a functional Y4 receptor response, and determining those drugs which inhibit the activation of the Y4 receptor in the cell using a bioassay, such as a second messenger assay, thereby identifying drugs which 10 act as Y4 receptor antagonists.

In one embodiment of the above-identified methods, the Y4 receptor is a human Y4 receptor. In another embodiment, the Y4 receptor is a rat Y4 receptor. In one embodiment, 15 the cell is a mammalian cell. In another embodiment, the mammalian cell is non-neuronal in origin. In a further embodiment, the mammalian cell non-neuronal in origin is a Cos-7 cell, a CHO cell, an LM(tk-) cell, a Y1 murine adrenal cell, or an NIH-3T3 cell.

20 The nucleic acid in the cell may have a coding sequence substantially the same as the coding sequences shown in Figures 1 and 3. Drug candidates are identified by choosing chemical compounds which bind with high affinity 25 to the expressed Y4 receptor protein in transfected cells, using radioligand binding methods well known in the art, examples of which are shown in the binding assays described herein. Drug candidates are also screened for selectivity by identifying compounds which 30 bind with high affinity to the Y4 receptor but do not bind with high affinity to any other NPY receptor subtype or to any other known receptor site. Because selective, high affinity compounds interact primarily with the target Y4 receptor site after administration to the 35 patient, the chances of producing a drug with unwanted side effects are minimized by this approach.

This invention provides a pharmaceutical composition comprising a drug identified by the method described above and a pharmaceutically acceptable carrier. As used herein, the term "pharmaceutically acceptable carrier" 5 encompasses any of the standard pharmaceutical carriers, such as a phosphate buffered saline solution, water, and emulsions, such as an oil/water or water/oil emulsion, and various types of wetting agents. Once the candidate drug has been shown to be adequately bio-available 10 following a particular route of administration, for example orally or by injection (adequate therapeutic concentrations must be maintained at the site of action for an adequate period to gain the desired therapeutic benefit), and has been shown to be non-toxic and 15 therapeutically effective in appropriate disease models, the drug may be administered to patients by that route of administration determined to make the drug bio-available, in an appropriate solid or solution formulation, to gain the desired therapeutic benefit.

20 This invention provides a nucleic acid probe comprising a nucleic acid molecule of at least 15 nucleotides capable of specifically hybridizing with a unique sequence included within the coding sequence of a nucleic acid molecule encoding a Y4 receptor, for example with a 25 coding sequence included within the sequences shown in Figures 1 and 3. In one embodiment, the nucleic acid encodes a human Y4 receptor. In another embodiment, the nucleic acid encodes a rat Y4 receptor. As used herein, 30 the phrase "specifically hybridizing" means the ability of a nucleic acid molecule to recognize a nucleic acid sequence complementary to its own and to form double-helical segments through hydrogen bonding between complementary base pairs. As used herein, a "unique 35 sequence" is a sequence specific to only the nucleic acid molecules encoding a Y4 receptor. Nucleic acid probe technology is well known to those skilled in the art who

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... will readily appreciate that such probes may vary greatly in length and may be labeled with a detectable label, such as a radioisotope or fluorescent dye, to facilitate detection of the probe. Detection of nucleic acid 5 encoding human Y4 receptors is useful as a diagnostic test for any disease process in which levels of expression of the corresponding Y4 receptor is altered. Nucleic acid probe molecules are produced by insertion of a nucleic acid molecule which encodes a Y4 receptor or 10 fragments thereof into suitable vectors, such as plasmids or bacteriophages, followed by insertion into suitable bacterial host cells and replication and harvesting of the nucleic acid probes, all using methods well known in the art. For example, the nucleic acid may be extracted 15 from a cell lysate using phenol and ethanol, digested with restriction enzymes corresponding to the insertion sites of the nucleic acid into the vector (discussed above), electrophoresed, and cut out of the resulting gel. Example of such nucleic acid molecules are shown in 20 Figures 1 and 3. The probes are useful for 'in situ' hybridization or in order to locate tissues which express this gene family, or for other hybridization assays for the presence of these genes or their mRNA in various biological tissues. In addition, synthesized 25 oligonucleotides (produced by a DNA synthesizer) complementary to the sequence of a DNA molecule which encodes a Y4 receptor or are useful as probes for these genes, for their associated mRNA, or for the isolation of related genes by homology screening of genomic or cDNA 30 libraries, or by the use of amplification techniques such as the Polymerase Chain Reaction. Synthesized oligonucleotides as described may also be used to determine the cellular localization of the mRNA produced by the Y4 gene by in situ hybridization.

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This invention also provides a method of detecting expression of a Y4 receptor by detecting the presence of

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mRNA coding for a Y4 receptor which comprises obtaining total mRNA from the cell using methods well known in the art and contacting the mRNA so obtained with a nucleic acid probe comprising a nucleic acid molecule of at least 5 15 nucleotides capable of specifically hybridizing with a sequence included within the sequence of a nucleic acid molecule encoding a Y4 receptor under hybridizing conditions, and detecting the presence of mRNA hybridized to the probe, thereby detecting the expression of the Y4 10 receptor by the cell. Hybridization of probes to target nucleic acid molecules such as mRNA molecules employs techniques well known in the art. In one possible means of performing this method, nucleic acids are extracted by precipitation from lysed cells and the mRNA is isolated 15 from the extract using a column which binds the poly-A tails of the mRNA molecules. The mRNA is then exposed to radioactively labelled probe on a nitrocellulose membrane, and the probe hybridizes to and thereby labels complementary mRNA sequences. Binding may be detected by 20 autoradiography or scintillation counting. However, other methods for performing these steps are well known to those skilled in the art, and the discussion above is merely an example.

25 This invention provides an antisense oligonucleotide having a sequence capable of specifically hybridizing with any sequences of an mRNA molecule which encodes a Y4 receptor so as to prevent translation of the mRNA molecule. The antisense oligonucleotide may have a 30 sequence capable of specifically hybridizing with any sequences of the cDNA molecule whose sequence is shown in Figure 1 or Figure 3. A particular example of an antisense oligonucleotide is an antisense oligonucleotide comprising chemical analogues of nucleotides.

35 This invention also provides a pharmaceutical composition comprising an amount of the oligonucleotide described

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... - above effective to reduce activity of a human Y4 receptor by passing through a cell membrane and specifically binding with mRNA encoding a Y4 receptor in the cell so as to prevent its translation and a pharmaceutically acceptable carrier capable of passing through a cell membrane. The oligonucleotide may be coupled to a substance which inactivates mRNA, such as a ribozyme. The pharmaceutically acceptable carrier capable of passing through cell membranes may also comprise a structure which binds to a receptor on a cell capable of being taken up by cells after binding to the structure. The structure of the pharmaceutically acceptable carrier may be capable of binding to a receptor which is specific for a selected cell type. The structure may be part of a protein known to bind a cell-type specific receptor, for example an insulin molecule, which would target pancreatic cells. Nucleic molecules having coding sequences substantially the same as the coding sequences shown in Figures 1 and 3 may be used as the oligonucleotides of the pharmaceutical composition.

This invention also provides a method of treating an abnormality wherein the abnormality is alleviated by decreasing the activity of a Y4 receptor which comprises administering to a subject an amount of the pharmaceutical composition described above effective to decrease the activity of the Y4 receptor. Several examples of such abnormal conditions are amnesia, anxiety, epilepsy, pain, hypertension, locomotor problems, circadian rhythm disorders, eating/body weight disorders, sexual/reproductive disorders, nasal congestion, diarrhea, gastrointestinal and cardiovascular disorders, and sleep and eating disorders.

35 Antisense oligonucleotide drugs inhibit translation of mRNA encoding these receptors. Synthetic oligonucleotides, or other antisense chemical structures

are designed to bind to mRNA encoding the Y4 receptor and inhibit translation of mRNA and are useful as drugs to inhibit expression of Y4 receptor genes in patients. This invention provides a means to therapeutically alter 5 levels of expression of human Y4 receptors by the use of a synthetic antisense oligonucleotide drug (SAOD) which inhibits translation of mRNA encoding these receptors. Synthetic oligonucleotides, or other antisense chemical structures designed to recognize and selectively bind to 10 mRNA, are constructed to be complementary to portions of the nucleotide sequences shown in Figures 1 and 3 of DNA, RNA or of chemically modified, artificial nucleic acids. The SAOD is designed to be stable in the blood stream for administration to patients by injection, or in laboratory 15 cell culture conditions, for administration to cells removed from the patient. The SAOD is designed to be capable of passing through cell membranes in order to enter the cytoplasm of the cell by virtue of physical and chemical properties of the SAOD which render it capable 20 of passing through cell membranes (e.g. by designing small, hydrophobic SAOD chemical structures) or by virtue of specific transport systems in the cell which recognize and transport the SAOD into the cell. In addition, the SAOD can be designed for administration only to certain 25 selected cell populations by targeting the SAOD to be recognized by specific cellular uptake mechanisms which binds and takes up the SAOD only within certain selected cell populations. For example, the SAOD may be designed to bind to a receptor found only in a certain cell type, 30 as discussed above. The SAOD is also designed to recognize and selectively bind to the target mRNA sequence, which may correspond to a sequence contained within the sequences shown in Figures 1 and 3 by virtue of complementary base pairing to the mRNA. Finally, the 35 SAOD is designed to inactivate the target mRNA sequence by any of three mechanisms: 1) by binding to the target mRNA and thus inducing degradation of the mRNA by

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intrinsic cellular mechanisms such as RNase I digestion, 2) by inhibiting translation of the mRNA target by interfering with the binding of translation-regulating factors or of ribosomes, or 3) by inclusion of other 5 chemical structures, such as ribozyme sequences or reactive chemical groups, which either degrade or chemically modify the target mRNA. Synthetic antisense oligonucleotide drugs have been shown to be capable of the properties described above when directed against mRNA 10 targets (J.S. Cohen, Trends in Pharm. Sci. 10, 435 (1989); H.M. Weintraub, Sci. Am. January (1990) p. 40). In addition, coupling of ribozymes to antisense oligonucleotides is a promising strategy for inactivating target mRNA (N. Sarver et al., Science 247, 1222 (1990)). 15 An SAOD serves as an effective therapeutic agent if it is designed to be administered to a patient by injection, or if the patient's target cells are removed, treated with the SAOD in the laboratory, and replaced in the patient. In this manner, an SAOD serves as a therapy to reduce 20 receptor expression in particular target cells of a patient, in any clinical condition which may benefit from reduced expression of Y4 receptors.

This invention provides an antibody directed to a Y4 25 receptor, for example a monoclonal antibody directed to an epitope of a Y4 receptor present on the surface of a cell and having an amino acid sequence substantially the same as an amino acid sequence for a cell surface epitope of the human Y4 receptor included in the amino acid 30 sequence shown in Figure 1 (Seq. I.D. No. 2) or the rat Y4 receptor included in the amino acid sequence shown in Figure 3 (Seq. I.D. No. 28). Amino acid sequences may be analyzed by methods well known in the art to determine whether they produce hydrophobic or hydrophilic regions 35 in the proteins which they build. In the case of cell membrane proteins, hydrophobic regions are well known to form the part of the protein that is inserted into the

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lipid bilayer which forms the cell membrane, while hydrophilic regions are located on the cell surface, in an aqueous environment. Therefore antibodies to the hydrophilic amino acid sequences shown in Figures 1 and 5 3 will probably bind to a surface epitope of a human or rat Y4 receptor, respectivley, as described. Antibodies directed to Y4 receptors may be serum-derived or monoclonal and are prepared using methods well known in the art. For example, monoclonal antibodies are prepared 10 using hybridoma technology by fusing antibody producing B cells from immunized animals with myeloma cells and selecting the resulting hybridoma cell line producing the desired antibody. Cells such as COS-7 cells or LM(tk-) cells comprising DNA encoding the human Y4 receptor and 15 thereby expressing the human Y4 receptor may be used as immunogens to raise such an antibody. Alternatively, synthetic peptides may be prepared using commercially available machines and the amino acid sequences shown in Figures 1 and 3 (Seq. I.D. Nos. 2 and 28). As a still 20 further alternative, DNA, such as a cDNA or a fragment thereof, may be cloned and expressed and the resulting polypeptide recovered and used as an immunogen. These antibodies are useful to detect the presence of human Y4 receptors encoded by the isolated DNA, or to inhibit the 25 function of the receptors in living animals, in humans, or in biological tissues or fluids isolated from animals or humans.

This invention provides a pharmaceutical composition 30 which comprises an amount of an antibody directed to the human Y4 receptor effective to block binding of ligands to the Y4 receptor, and a pharmaceutically acceptable carrier. A monoclonal antibody directed to an epitope of a Y4 receptor present on the surface of a cell and having 35 an amino acid sequence substantially the same as an amino acid sequence for a cell surface epitope of the Y4 receptor included in the amino acid sequences shown in

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Figures 1 and 3 is useful for this purpose. In one embodiment, the Y4 receptor is a human Y4 receptor. In another embodiment, the Y4 receptor is a rat Y4 receptor.

5 This invention also provides a method of treating an abnormality wherein the abnormality is alleviated by decreasing the activity of a Y4 receptor which comprises administering to a subject an amount of the pharmaceutical composition described above effective to
10 block binding of ligands to the Y4 receptor, thereby treating the abnormality. Binding of the antibody to the receptor prevents the receptor from functioning, thereby neutralizing the effects of Y4 receptor activity. The monoclonal antibodies described above are both useful for
15 this purpose. Some examples of abnormalities are amnesia, depression, anxiety, epilepsy, pain, depression, hypertension, and sleep and eating disorders.

This invention provides a method of detecting the
20 presence of a Y4 receptor on the surface of a cell which comprises contacting the cell with an antibody directed to the Y4 receptor, under conditions permitting binding of the antibody to the receptor, and detecting the presence of the antibody bound to the cell, thereby
25 detecting the presence of the Y4 receptor on the surface of the cell. Such a method is useful for determining whether a given cell is defective in activity of Y4 receptors on the surface of the cell. Bound antibodies are detected by methods well known in the art, for
30 example by binding fluorescent markers to the antibodies and examining the cell sample under a fluorescence microscope to detect fluorescence on a cell indicative of antibody binding. The monoclonal antibodies described above are useful for this purpose.

35

This invention provides a transgenic nonhuman mammal expressing nucleic acid encoding a Y4 receptor. This

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invention also provides a transgenic nonhuman mammal comprising a homologous recombination knockout of the native Y4 receptor. This invention also provides a transgenic nonhuman mammal whose genome comprises 5 antisense DNA complementary to DNA encoding a Y4 receptor so placed as to be transcribed into antisense mRNA which is complementary to mRNA encoding a Y4 receptor and which hybridizes to mRNA encoding a Y4 receptor thereby reducing its translation. The DNA may additionally 10 comprise an inducible promoter or additionally comprise tissue specific regulatory elements, so that expression can be induced, or restricted to specific cell types. Examples of DNA are DNA or cDNA molecules having a coding sequence substantially the same as the coding sequences 15 shown in Figures 1 and 3. An example of a transgenic animal is a transgenic mouse. Examples of tissue specificity-determining regions are the metallothionein promotor (Low, M.J., Lechan, R.M., Hammer, R.E. et al. Science 231:1002-1004 (1986)) and the L7 promotor 20 (Oberdick, J., Smeyne, R.J., Mann, J.R., Jackson, S. and Morgan, J.I. Science 248:223-226 (1990)).

Animal model systems which elucidate the physiological and behavioral roles of Y4 receptors are produced by 25 creating transgenic animals in which the activity of a Y4 receptor is either increased or decreased, or the amino acid sequence of the expressed Y4 receptor protein is altered, by a variety of techniques. Examples of these techniques include: 1) Insertion of normal or mutant 30 versions of DNA encoding a Y4 receptor or homologous animal versions of these genes, by microinjection, retroviral infection or other means well known to those skilled in the art, into appropriate fertilized embryos in order to produce a transgenic animal (Hogan B. et al. 35 Manipulating the Mouse Embryo, A Laboratory Manual, Cold Spring Harbor Laboratory (1986)). 2) Homologous recombination (Capecchi M.R. Science 244:1288-1292

(1989); Zimmer, A. and Gruss, P. *Nature* 338:150-153 (1989)) of mutant or normal, human or animal versions of these genes with the native gene locus in transgenic animals to alter the regulation of expression or the 5 structure of these Y4 receptors. The technique of homologous recombination is well known in the art. It replaces the native gene with the inserted gene and so is useful for producing an animal that cannot express native receptor but does express, for example, an inserted 10 mutant receptor, which has replaced the native receptor in the animal's genome by recombination, resulting in underexpression of the receptor. Microinjection adds genes to the genome, but does not remove them, and so is useful for producing an animal which expresses its own 15 and added receptors, resulting in overexpression of the receptor. One means available for producing a transgenic animal, with a mouse as an example, is as follows: Female mice are mated, and the resulting fertilized eggs are dissected out of their oviducts. The eggs are stored 20 in an appropriate medium such as M2 medium (Hogan B. et al. *Manipulating the Mouse Embryo, A Laboratory Manual*, Cold Spring Harbor Laboratory (1986)). DNA or cDNA encoding a human Y4 receptor is purified from a vector (such as plasmid pcEXV-Y4 described above) by methods 25 well known in the art. Inducible promoters may be fused with the coding region of the DNA to provide an experimental means to regulate expression of the trans-gene. Alternatively or in addition, tissue specific regulatory elements may be fused with the coding region 30 to permit tissue-specific expression of the trans-gene. The DNA, in an appropriately buffered solution, is put into a microinjection needle (which may be made from capillary tubing using a pipet puller) and the egg to be injected is put in a depression slide. The needle is 35 inserted into the pronucleus of the egg, and the DNA solution is injected. The injected egg is then transferred into the oviduct of a pseudopregnant mouse (a

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mouse stimulated by the appropriate hormones to maintain pregnancy but which is not actually pregnant), where it proceeds to the uterus, implants, and develops to term.

As noted above, microinjection is not the only method 5 for inserting DNA into the egg cell, and is used here only for exemplary purposes.

Since the normal action of receptor-specific drugs is to activate or to inhibit receptor activity, the transgenic 10 animal model systems described above are useful for testing the biological activity of drugs directed against these Y4 receptors even before such drugs become available. These animal model systems are useful for predicting or evaluating possible therapeutic 15 applications of drugs which activate or inhibit Y4 receptor activity by inducing or inhibiting expression of the native or trans-gene and thus increasing or decreasing activity of normal or mutant Y4 receptors in the living animal. Thus, a model system is produced in 20 which the biological activity of drugs directed against these Y4 receptors are evaluated before such drugs become available. The transgenic animals which have increased or decreased Y4 receptor activity indicate by their physiological state whether increase or decrease of the 25 Y4 receptor activity is therapeutically useful. It is therefore useful to evaluate drug action based on the transgenic model system. One use is based on the fact that it is well known in the art that a drug such as an antidepressant acts by blocking neurotransmitter uptake, 30 and thereby increases the amount of neurotransmitter in the synaptic cleft. The physiological result of this action is to stimulate the production of less receptor by the affected cells, leading eventually to decreased activity of the receptor. Therefore, an animal which has 35 decreased receptor activity is useful as a test system to investigate whether the actions of such drugs which result in decreased receptor activity are in fact

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therapeutic. Another use is that if increased receptor activity is found to lead to abnormalities, then a drug which down-regulates or acts as an antagonist to Y4 receptor is indicated as worth developing, and if a 5 promising therapeutic application is uncovered by these animal model systems, activation or inhibition of Y4 receptor activity is achieved therapeutically either by producing agonist or antagonist drugs directed against these Y4 receptors or by any method which increases or 10 decreases the activity of these Y4 receptors.

This invention provides a method of determining the physiological effects of expressing varying levels of human Y4 receptors which comprises producing a transgenic 15 nonhuman animal whose levels of human Y4 receptor activity are varied by use of an inducible promoter which regulates Y4 receptor expression. This invention also provides a method of determining the physiological effects of expressing varying levels of Y4 receptors 20 which comprises producing a panel of transgenic nonhuman animals each expressing a different amount of Y4 receptor activity. Such animals may be produced by introducing different amounts of nucleic acid encoding a Y4 receptor into the oocytes from which the transgenic animals are 25 developed.

This invention also provides a method for identifying a Y4 receptor antagonist capable of alleviating an abnormality in a subject, wherein the abnormality is 30 alleviated by decreasing the activity of a Y4 receptor which comprises administering the antagonist to a transgenic nonhuman mammal expressing at least one artificially introduced nucleic acid molecule encoding a Y4 receptor and determining whether the antagonist 35 alleviates the physical and behavioral abnormalities displayed by the transgenic nonhuman mammal as a result of activity of a Y4 receptor, thereby identifying a Y4

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receptor antagonist. As used herein, the term "antagonist" means a compound or composition which may be natural, synthetic, or a product derived from screening. Examples of nucleic acid molecules are DNA, cDNA, genomic

5 DNA, synthetic DNA or RNA molecules having coding sequences substantially the same as the coding sequences shown in Figures 1 and 3. This invention also provides an antagonist identified by the method described above.

10 This invention provides a pharmaceutical composition comprising an amount of the antagonist described supra and a pharmaceutically acceptable carrier.

15 This invention further provides a method for treating an abnormality in a subject wherein the abnormality is alleviated by decreasing the activity of a Y4 receptor which comprises administering to the subject an effective amount of the pharmaceutical composition described above, thereby treating the abnormality.

20 This invention provides a method for identifying a Y4 receptor agonist capable of alleviating an abnormality in a subject wherein the abnormality is alleviated by activation of a Y4 receptor which comprises administering

25 the agonist to the transgenic nonhuman mammals described above determining whether the agonist alleviates the physical and behavioral abnormalities displayed by the transgenic nonhuman mammal, the alleviation of the abnormality indicating the identification of a Y4

30 receptor agonist.

This invention provides an agonist identified by the method described above.

35 This invention also provides a pharmaceutical composition comprising an amount of the agonist identified by the method described above and a pharmaceutically acceptable

carrier.

This invention further provides a method for treating an abnormality in a subject wherein the abnormality is
5 alleviated by activation of a Y4 receptor which comprises administering to a subject an effective amount of the pharmaceutical composition described above, thereby treating the abnormality.

- 10 This invention provides a method for diagnosing a predisposition to a disorder associated with the activity of a specific Y4 receptor allele which comprises: a) obtaining DNA of subjects suffering from the disorder; b) performing a restriction digest of the DNA with a panel
15 of restriction enzymes; c. electrophoretically separating the resulting DNA fragments on a sizing gel; d) contacting the resulting gel with a nucleic acid probe capable of specifically hybridizing to DNA encoding a Y4 receptor and labelled with a detectable marker; e)
20 detecting labelled bands which have hybridized to the DNA encoding a Y4 receptor labelled with a detectable marker to create a unique band pattern specific to the DNA of subjects suffering from the disorder; f) preparing DNA obtained for diagnosis by steps a-e; and g) comparing
25 the unique band pattern specific to the DNA of subjects suffering from the disorder from step e and the DNA obtained for diagnosis from step f to determine whether the patterns are the same or different and thereby to diagnose predisposition to the disorder if the patterns
30 are the same. This method may also be used to diagnose a disorder associated with the expression of a specific Y4 receptor allele.

This invention provides a method of preparing the purified, isolated Y4 receptor which comprises a)
35 constructing a vector adapted for expression in a cell which comprises the regulatory elements necessary for the

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expression of nucleic acid in the cell operatively linked to the nucleic acid encoding a Y4 receptor as to permit expression thereof, wherein the cell is selected from the group consisting of bacterial cells, yeast cells, insect 5 cells and mammalian cells; b) inserting the vector of step a in a suitable host cell; c) incubating the cells of step b under conditions allowing the expression of a Y4 receptor; d) recovering the receptor so produced; and e) purifying the receptor so recovered, thereby preparing 10 the purified, isolated Y4 receptor. An example of an isolated Y4 receptor is an isolated protein having substantially the same amino acid sequence as the amino acid sequences shown in Figures 1 and 3. For example, cells can be induced to express receptors by exposure to 15 substances such as hormones. The cells can then be homogenized and the receptor isolated from the homogenate using an affinity column comprising, for example, PP or another substance which is known to bind to the receptor. The resulting fractions can then be purified by 20 contacting them with an ion exchange column, and determining which fraction contains receptor activity or binds anti-receptor antibodies. This method for preparing Y4 receptor uses recombinant DNA technology methods well known in the art. For example, isolated 25 nucleic acid encoding Y4 receptor is inserted in a suitable vector, such as an expression vector. A suitable host cell, such as a bacterial cell, or a eukaryotic cell such as a yeast cell, is transfected with the vector. Y4 receptor is isolated from the culture 30 medium by affinity purification or by chromatography or by other methods well known in the art.

This invention identifies for the first time a new receptor protein, its amino acid sequence, and its human 35 gene. Furthermore, this invention describes a previously unrecognized group of receptors within the definition of a Y4 receptor. The information and experimental tools

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provided by this discovery are useful to generate new therapeutic agents, and new therapeutic or diagnostic assays for this new receptor protein, its associated mRNA molecule or its associated genomic DNA. The information and experimental tools provided by this discovery will be useful to generate new therapeutic agents, and new therapeutic or diagnostic assays for this new receptor protein, its associated mRNA molecule, or its associated genomic DNA.

10

Specifically, this invention relates to the first isolation of a human and a rat genomic clone encoding a Y4 receptor. A new human gene for the receptor identified herein as Y4 has been identified and characterized. In addition, the human Y4 receptor has been expressed in COS-7 cells. The pharmacological binding properties of the protein encoded have been determined, and these binding properties classify this protein as a novel NPY/PYY/PP receptor which we designate as a Y4 receptor. Mammalian cell lines expressing this Y4 receptor at the cell surface have been constructed, thus establishing the first well-defined, cultured cell lines with which to study this Y4 receptor.

25 This invention will be better understood by reference to the Experimental Details which follow, but those skilled in the art will readily appreciate that the specific experiments detailed are only illustrative of the invention as described more fully in the claims which 30 follow thereafter.

Experimental Details

Cloning and Sequencing of a human (Y4) Neuropeptide Receptor. A human placenta genomic library in λ dash II ($\sim 1.5 \times 10^6$ total recombinants; Stratagene, LaJolla, CA) was screened using overlapping transmembrane (TM) oligonucleotide probes (TM 1, 2, 3, 5 and 7) derived from the rat Y1 neuropeptide receptor gene (Eva, C. et al., 1990; GenBank accession No. Z11504). Overlapping oligomers (TM1: nts. 198-251, (+)strand/5'-TTGCTTATGGGGCTGTGATTATTCTTGGGGTCTCTGGAAACCTGG-3' (Sequence I.D. No. 3) and (-)strand/5'-TAGGATGATTATGATCAATGCCAGGTTCCAGAGACCCCAAGAAT-3' (Sequence I.D. No. 4); TM2: nts. 269-328, (+)strand/5'-AAAGAGATGAGGAATGTCACCAACATTCTGATCGTGAACCTCTCC-3' (Sequence I.D. No. 5) and (-)strand/5'-CAGCAAGTCTGAGAAGGAGAGGTTCACGATCAGAATGTTGGTGAC-3' (Sequence I.D. No. 6); TM3: nts. 401-478, (+)strand/5'-TGCAAAC TGAAATCCTTTGTGCAATGCGTCTCCATTACAGTATCCATTCTCT-3' (Sequence I.D. No. 7) and (-) strand/5'-ACGTTCCACAGCGATGAGAACCAGAGAGAAAATGGATACTGTAATGGAGACGCA-3' (Sequence I.D. No. 8); TM5: nts. 716-778, (+)strand/5'-CTGCAGTATTTGGCCCACCTCTGTTCATATTGATATGCTAC-3' (Sequence I.D. No. 9) and (-) strand/5'-CAAGCGAATGTATATCTTGAAGTAGCATATGAATATGAAACA-3' (Sequence I.D. No. 10); TM7: nts. 971-1045, (+)strand/5'-CTGCTCTGCCACCTCACGGCCATGATCTCCACCTGCGTCAACC-3' (Sequence I.D. No. 11) and (-)strand/5'-GAAATTTTGTTCAGGAATCCATAAAAGATGGGGTTGA CGCAGGTGGA-3' (Sequence I.D. No. 12); GenBank accession No. Z11504) were labeled with [32 P]dATP and [32 P]dCTP by synthesis with the large fragment of DNA polymerase. Hybridization was performed at low stringency conditions: 40°C. in a solution containing 25.0% formamide, 5x SSC (1X SSC is 0.15M sodium chloride, 0.015M sodium citrate), 1x Denhardt's solution (0.02% polyvinylpyrrolidone,

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0.02% Ficoll, 0.02% bovine serum albumin), and 25 μ g/ μ l sonicated salmon sperm DNA. The filters were washed at 40°C. in 0.1x SSC containing 0.1% sodium dodecyl sulfate and exposed at -70°C. to Kodak XAR film in the presence 5 of an intensifying screen. Lambda phage clones hybridizing with the probes were plaque purified and DNA was prepared for Southern blot analysis (Southern, 1975; Sambrook et al., 1989). A Genomic clone hybridizing with all five of the rat Y1 TM probes, designated hp25a, 10 was isolated using this method. For subcloning and further Southern blot analysis, the hp25a DNA was cloned into pUC18 (Pharmacia, Piscataway, NJ). Nucleotide sequence analysis was accomplished by the Sanger dideoxy 15 nucleotide chain termination method (Sanger et al., 1977) on denatured double-stranded plasmid templates, using Sequenase (US Biochemical Corp., Cleveland, OH).

Cloning and Sequencing of a rat NPY (Y4) neuropeptide receptor:

20 A rat spleen genomic library (Stratagene, La Jolla, CA) was screened using overlapping TM oligonucleotide probes (TM 1 - 7) derived from the nucleotide sequences corresponding approximately to the TM regions of the amino acid sequence of the human Y4 receptor as shown in 25 Figure 2. The overlapping oligomers used were as follows:

TM1: nts. #129-201,

(+) strand/5'-TCATCGTCACCTCCTACAGCATTGAGACTGTCGTGG
GGGTCCCTGGGT (Seq. ID. No. 13) and

(-) strand/5'-ACAGTCACACACATCAGGCAGAGGTTACCCAGGAC
CCCCACGACAG (Seq. I.D. No. 14);

TM2: nts. #234-303,

(+) strand/5'-TGCTTATCGCCAACCTGGCCTCTCTGACTTCCTCATGTGCCTCC (Seq.
ID. No. 15) and

(-) strand/5'-TAGACGGCGGTCAAGCGGCTGGCAGAGGAGGCACATGAGGAAGTCA (Seq.
ID. No. 16);

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TM3: nts. #348-417,

(+) s t r a n d / 5'

TGTCGGCCTTCATCCAGTGCATGTCGGTACGGTCTCCATCCTCT ~~Seq.~~ IDNo. 17
I.D. No.) and

5 (-) s t r a n d / 5'

CTCTCCAGGGCCACGAGGACGAGCGAGAGGATGGAGACCGTCACC ~~Seq.~~ IDNo. 18
I.D. No.);

TM4: nts. #467-536,

(+) s t r a n d / 5'

10 GCCTACCTGGGGATTGTGCTCATCTGGGTATTGCCTGTGTCCTC ~~Seq.~~ IDNo. 19
I.D. No.) and

(-) s t r a n d / 5'

TGCTGTTGCCAGGAAGGGCAGGGAGAGGACACAGGAATGACCC ~~Seq.~~ IDNo. 20
I.D. No.);

15 TM5: nts. #637-706,

(+) s t r a n d / 5'

CATCTACACCACCTTCCTGCTCCTCTTCCAGTACTGCCTCCACT ~~Seq.~~ IDNo. 21
I.D. No.) and

(-) s t r a n d / 5'

20 TGCATAACAGACCAGGATGAAGCCCAGTGGGAGGCAGTACTGGAA ~~Seq.~~ IDID No. 22
I.D. No.);

TM6: nts. #800-870,

(+) strand/5' -CTGGTGGTGATGGTGGTGGCCTTGCCGTGCTCT

~~Seq.~~ ID No. 23
GGCTGCCTCTGC (Seq. I.D. No.) and

25 (-) s t r a n d / 5'

CAGTCTCCAGGCTGTTAACACATGCAGAGGCAGCCAGAGCACG ~~Seq.~~ IDNo. 24
I.D. No.);

TM7: NTS. #908-977,

(+) s t r a n d / 5'

30 ATCTTCTTAGTGTGCCACTTGCTTGCCATGGCCTCCACCTGCGTC ~~Seq.~~ IDNo. 25
I.D. No.) and

(-) s t r a n d / 5'

TGAGAAAGCCATAGATGAATGGGTTGACGCAGGTGGAGGCCATGG ~~Seq.~~ IDNo. 26
I.D. No.) were labeled with [³²P]-ATP and [³²P]-CTP

35 by synthesis with the large fragment of DNA polymerase. Hybridization was performed at reduced stringency conditions: 40°C in a solution

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containing 37.5% formamide, 10% dextran sulfate, 5X
SSC, 1X Denhardt's solution, and 100 µg/ml of
sonicated salmon sperm DNA. The filters were washed
5 at 45°C in 0.1X SSC containing 0.1% sodium dodecyl
sulfate (SDS) and exposed at -70°C to Kodak XAR film
in the presence of an intensifying screen. Lambda
phage clones hybridizing to the probes were plaque
purified by successive plating and rescreening. A
10 genomic clone hybridizing with all seven human Y4
receptor TM probes, designated rs16b, was isolated
using this method. For expression and sequence
analysis, a 2.0 kb BamHI/HindIII fragment of rs16b
was subcloned into the corresponding polylinker
sites of a pcEXV-3 eukaryotic expression vector
15 (Miller and Germain, 1986) modified to include a
polylinker with EcoRI, SstI, ClaI, KpnI, SmaI, XbaI,
BamHI, SalI and HindIII restriction sites and
designated EXJ.RH. Nucleotide sequence analysis was
accomplished by the Sanger dideoxy nucleotide chain-
20 termination method (Sanger, 1977) on double stranded
plasmid templates, using Sequenase (U.S. Biochemical
Corp., Cleveland, Ohio).

Transient Transfection: The entire coding region of
25 hp25a (1127 bp), including 680 bp of 5' untranslated (5'
UT) and 205 bp of 3' untranslated sequence (3' UT), was
cloned into the BamHI and EcoRI sites of the polylinker-
modified eukaryotic expression vector pCEXV-3 (Miller et
al., 1986), called EXJ.HR (J.B., unpublished data).
30 Monkey kidney cells (Cos-7) were transiently transfected
with plasmid hp25a/EXJ (expression vector containing the
hp25a receptor gene) using DEAE dextran methodology
(reagents obtained from Specialty Media, Lavellette, NJ).

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the rs16b receptor gene), was transiently transfected into Cos-7 cells using similar methods, as were the human Y1 receptor (Larhammar, 1992) and the human Y2 receptor. The cloned Y2 receptor was disclosed in U.S. patent 5 application 08/192,288 filed on February 2, 1994, currently pending, the foregoing contents of which are hereby incorporated by reference.

Stable Transfection

10 Human Y4 receptors were co-transfected with a G-418 resistant gene into the mouse embryonic NIH-3T3 cell line by a calcium phosphate transfection method (Cullen, 1987). Stably transfected cells were selected with G-418. Human Y4 receptors were similarly transfected into 15 mouse fibroblast LM(tk-) cells.

Cell culture: COS-7 cells were grown on 150 mm plates (Corning) in D-MEM with supplements (Dulbecco's Modified Eagle Medium with 10% bovine calf serum, 2 mM glutamine, 20 100 units/ml penicillin/80 units/ml streptomycin) at 37 °C, 5% CO₂. Stock plates of COS-7 cells were trypsinized and split 1:6 every 3-4 days. SK-N-Be(2) human neuroblastoma cells were grown similarly in 225 cm² flasks (Co-star) using 50% Eagle's Modified Essential Media, 50% 25 Ham's Nutrient Mixture F-12, 15% fetal bovine serum, 2 mM glutamine, 100 units/ml penicillin/80 units/ml streptomycin, and 1% non-essential amino acids. Stock flasks of SK-N-Be(2) cells were trypsinized and split 1:10 every 7 days.

30

Mouse embryonic NIH-3T3 cells were grown on 150 mm plates in Dulbecco's Modified Eagle Medium (DMEM) with supplements (10% bovine calf serum, 4 mM glutamine, 100 units/ml penicillin/100 µg/ml streptomycin) at 37 °C, 5% CO₂. Stock plates of NIH-3T3 cells were trypsinized and split 1:15 every 3-4 days. Mouse fibroblast LM(tk-)

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cells were grown on 150 mm plates in Dulbecco's Modified Eagle Medium (DMEM) with supplements (10% bovine calf serum, 4 mM glutamine, 100 units/ml penicillin/100 µg/ml streptomycin) at 37 °C, 5% CO₂. Stock plates of LM(tk-) 5 cells were trypsinized and split 1:10 every 3-4 days.

Cell culture media and supplements were from Specialty Media (Lavallette, NJ). Cell culture plates (150 mm) were from Corning (Corning, NY). Cell culture flasks (225 cm²) 10 and polypropylene microtiter plates were from Co-star (Cambridge, MA).

Membrane Harvest: Membranes were harvested from COS-7 cells 48 hours after transfection and from SK-N-Be(2) 15 seven days after splitting. Adherent cells were washed twice in ice-cold phosphate buffered saline (138 mM NaCl, 8.1 mM Na₂HPO₄, 2.5 mM KCl, 1.2 mM KH₂PO₄, 0.9 mM CaCl₂, 0.5 mM MgCl₂, pH 7.4) and lysed by sonication in ice-cold 20 hypotonic buffer (20 mM Tris-HCl, 5 mM EDTA, pH 7.7). Large particles and debris were cleared by low speed centrifugation (200 x g, 10 min, 4 °C). Membranes were 25 collected from the supernatant fraction by high speed centrifugation (32,000 x g, 18 min, 4 °C), washed with ice-cold hypotonic buffer, and collected again by high speed centrifugation (32,000 x g, 18 min, 4 °C). The final membrane pellet was resuspended by sonication into a small volume (~500 µl) of ice-cold binding buffer (10 mM NaCl, 20 mM HEPES, 0.22 mM KH₂PO₄, 1.26 mM CaCl₂, 0.81 mM MgSO₄, pH 7.4). Protein concentration was measured by the 30 Bradford method (Bradford, 1976) using Bio-Rad Reagent, with bovine serum albumin as a standard.

Radioligand Binding to Membrane Suspensions: Membrane suspensions were diluted in binding buffer supplemented 35 with 0.1% bovine serum albumin and 0.1% bacitracin to yield an optimal membrane protein concentration: ~ 0.02 mg/ml for human Y1 receptors, ~ 0.015 mg/ml for hp25a

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receptors, and - 0.25 mg/ml for SK-N-Be(2). (Under these conditions, ^{125}I -PYY bound by membranes in the assay was less than 10% of ^{125}I -PYY delivered to the sample.) ^{125}I -PYY and non-labeled peptide competitors were also diluted to 5 desired concentrations in supplemented binding buffer. Individual samples were then prepared in 96-well polypropylene microtiter plates by mixing membrane suspensions (200 μl), ^{125}I -PYY (25 μl), and non-labeled peptides or supplemented binding buffer (25 μl).

10 Samples were incubated in a 30 °C water bath with constant shaking for 120 min. Incubations were terminated by filtration over Whatman GF/C filters (pre-coated with 0.5% polyethyleneimine and air-dried before use). Filter-trapped membranes were counted for ^{125}I in a gamma counter.

15 Non-specific binding was defined by 100 nM human PP for hp25a receptors and by 100 nM NPY for Y1 and SK-N-Be(2) receptors. Specific binding in time course and competition studies was typically 80%; most non-specific binding was associated with the filter. Binding data were 20 analyzed using nonlinear regression and statistical techniques available in the GraphPAD InPlot package (San Diego, CA).

Porcine ^{125}I -PYY was from New England Nuclear (Boston, MA). NPY and related peptide analogs were from either Bachem 25 California (Torrance, CA) or Peninsula (Belmont, CA). Whatman GF/C filters were from Brandel (Gaithersburg, MD). Bio-Rad Reagent was from Bio-Rad (Hercules, CA). Bovine serum albumin and bacitracin were from Sigma (St. Louis, MO). All other materials were reagent grade.

30 Functional Assay: Radioimmunoassay of cAMP
Stably transfected cells were seeded into 96-well microtiter plates and cultured until confluent. To reduce the potential for receptor desensitization, the serum component of the media was reduced to 1.5% for 4 to 16 35 hours before the assay. Cells were washed in Hank's buffered saline, or HBS (150 mM NaCl, 20 mM HEPES, 1 mM CaCl₂, 5 mM KCl, 1 mM MgCl₂, and 10 mM glucose)

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supplemented with 0.1% bovine serum albumin plus 3 mM theophylline and pre-equilibrated in the same solution for 20 min at 37 °C in 5% CO₂. Cells were then incubated 5 min with 10 µM forskolin and various concentrations of 5 receptor-selective ligands. The assay was terminated by the removal of HBS and acidification of the cells with 100 mM HCl. Intracellular cAMP was extracted and quantified with a modified version of a magnetic bead-based radioimmunoassay (Advanced Magnetics, Cambridge, MA). The final antigen/antibody complex was separated 10 from free ¹²⁵I-cAMP by vacuum filtration through a PVDF filter in a microtiter plate (Millipore, Bedford, MA). Filters were punched and counted for ¹²⁵I in a Packard gamma counter. Binding data were analyzed using nonlinear 15 regression and statistical techniques available in the GraphPAD Prism package (San Diego, CA).

Functional Assay: Intracellular Calcium Mobilization

The intracellular free calcium concentration was 20 measured by microspectrofluorometry using the fluorescent indicator dye Fura-2/AM. Stably transfected cells were seeded onto a 35 mm culture dish containing a glass coverslip insert. Cells were washed with HBS and then loaded with 100 µl of Fura-2/AM (10 µM) for 20 to 40 min. 25 After washing with HBS to remove the Fura-2/AM solution, cells were equilibrated in HBS for 10 to 20 min. Cells were then visualized under the 40X objective of a Leitz Fluovert FS microscope and fluorescence emission was determined at 510 nM with excitation wave lengths 30 alternating between 340 and 380 nM. Raw fluorescence data were converted to calcium concentrations using standard calcium concentration curves and software analysis techniques.

35 Tissue Localization and Gene Expression: Reverse Transcriptase PCR

Human tissues (obtained from National Disease Research

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interchange) were homogenized and total RNA extracted using guanidine isothiocyanate/CsCl cushion method (Kingston, 1987). RNA was treated with DNase to remove any contaminating genomic DNA. cDNA was prepared from

5 total RNA with random hexanucleotide primers using reverse transcriptase (Superscript II; BRL). An aliquot of the first strand cDNA (250ng of total RNA) was amplified in a 50 μ l PCR reaction mixture (200 μ M dNTPs final concentration) containing 1.2U of Taq polymerase

10 in the buffer supplied by the manufacturer (Perkin-Elmer Corporation), and 1 μ M of primers, using a program consisting of 30 cycles of 94°C./2', 68°C./2', and 72°C./3', with a pre- and post-incubation of 95°C./5' and 72°C./10', respectively. PCR primers for human Y4 were

15 designed against the human Y4 sequence in the third intracellular loop and carboxy terminal regions: 5'-
CGCGTGTTCACAAGGGCACCTA-3' and 5'-TGCCACTTAGCCTCAGGGACCC-
3', respectively.

20 The PCR products were run on a 1.5% agarose gel and transferred to charged nylon membranes (Zetaprobe GT, BioRad), and analyzed as Southern blots. Hybridization probes corresponding to the receptor region flanked by PCR primers were prepared (5' -

25 TCCGTATGTACTGTGGACAGGGGCAGATGCTCCGACTCCTCCAGG-3') and pre-screened for the absence of cross-reactivity with human Y1 and human Y2 receptor subtypes. Filters were hybridized with end-labeled [γ -³²P]ATP internal probe to the PCR primers, washed under high stringency, and

30 exposed to Kodak XAR film in the presence of an intensifying screen, as described above. Similar PCR and Southern blot analysis were conducted with primers and probe directed to the housekeeping gene, glyceraldehyde-3-phosphate dehydrogenase (Clontech, Palo

35 Alto, CA), and demonstrated that equal amounts of cDNA from the different tissues were being assayed for NPY expression.

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Results

A human genomic placenta library was screened, under 5 reduced stringency conditions, with oligonucleotide probes directed to the first, second, third, fifth, and seventh transmembrane regions of the rat Y1 neuropeptide receptor gene (Eva, C. et al., 1990; GenBank accession No. Z11504). Positively-hybridizing clones (~100-150) 10 were isolated, plaque-purified and characterized by Southern blot analysis and sequencing. One clone, hp25a, contained a 1.3 kb PstI fragment which hybridized with the rat Y1-derived oligonucleotide probes and was subsequently subcloned into a pUC vector. DNA sequence 15 analysis indicated greatest homology to the rat and human Y1 receptor genes. This clone was a partial intronless gene fragment, encoding part of the third intracellular loop through the carboxyl terminus, including a termination codon.

20 In order to obtain a full-length clone, a 2.0 kb BamHI/EcoRI hybridizing fragment, containing the entire coding region, which was intronless, was subcloned into an expression vector and sequenced. The genomic full-length construct in the expression vector (called hp25a/EXJ) contains an open reading frame of 1127 bp, with 680 bp of the predicted 5' UT and 205 bp of predicted 3' UT sequence, and encodes a protein of 375 aa in length, with a relative molecular mass of ~41,000 25 daltons. Hydropathy analysis of the protein is consistent with a putative topography of seven transmembrane domains, indicative of the G protein-coupled receptor family.

30 Initial sequence analysis revealed that clone hp25a/EXJ contained several conserved structural features/residues found among the members of the neuropeptide receptor

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family, including two glycines and asparagine in TM1 (positions 55, 58 and 59, respectively, in Fig. 2), an asparagine, leucine and aspartic acid in TM2 (positions 82, 83, and 87, respectively, in Fig. 2), a serine and 5 leucine in TM3 (positions 128 and 132, respectively, in Fig. 2), a tryptophan and proline in TM4 (positions 164 and 173, respectively, in Fig. 2), a tyrosine and proline in TM5 (positions 223 and 226, respectively, in Fig. 2), a phenylalanine, tryptophan, and proline in TM6 10 (positions 275, 279, and 281, respectively, in Fig. 2), and a serine, threonine, asparagine, and proline in TM7 (positions 315, 316, 319, and 320, respectively, in Fig. 2). Other features of this human hp25a receptor gene are 15 the presence of three potential sites for N-linked glycosylation in the amino terminus (asparagine residues 2, 19, and 29; Fig. 1) and the presence of several serines and threonines in the carboxyl terminus and intracellular loops, which may serve as sites for potential phosphorylation by protein kinases.

20 A comparison of nucleotide and peptide sequences of clone hp25a/EXJ with sequences contained in the Genbank/EMBL databases reveals that the clone is most related to the rat, mouse and human Y1 receptor genes and proteins (see 25 Fig. 2). The hp25a clone exhibits 42% overall amino acid identity with the human NPY-1 receptor and 55% identity when comparing only the transmembrane domains between hp25a and Y1. The comparison of the individual amino acid residues in the TM domains between hp25a and Y1 30 reveal <30%, 57%, 57%, 57%, 52%, 63%, and 71% identity in the corresponding one through seven TM regions, respectively. The hp25a clone hybridized only with the TM7-specific probe from the original set of rat-derived TM probes originally used to screen the library which is 35 consistent with the hp25a clone sharing the highest degree of amino acid identity with the TM7 domain of the rat Y1 receptor.

A rat homolog of the human Y4 receptor, designated rs16b, was isolated from a rat spleen genomic library using probes derived from the transmembrane regions of the 5 human Y4 receptor. The nucleotide sequence of rs16b is 80% identical in the coding region to the nucleotide sequence of the human Y4 receptor, and encodes a protein 375 amino acids in length (Figure 3). The rs16b clone exhibits 75% overall amino acid identity with the human 10 Y4 amino acid sequence, and in the putative transmembrane domains (TMs), the protein predicted by rs16b exhibits 84% amino acid identity with the human Y4 receptor. This degree of primary amino acid sequence identity is lower than is typically seen for species homologues, and 15 suggests that rat and human Y4 receptors may exhibit functional variations as well. The predicted intracellular loop between TMs V and VI is particularly divergent, showing only 56% amino acid identity between rat and human Y4; divergence in this region could 20 potentially mediate differences in G-protein coupling between the rat and human receptors. The primary sequences of rat and human Y4 receptors also show differences in their patterns of sequence motifs for casein kinase II phosphorylation, N-myristoylation, and 25 protein kinase C phosphorylation; these sites could potentially mediate differences in the function or regulation of the two receptors.

Monkey kidney cells transiently expressing the gene 30 encoding the hp25a receptor were used for pharmacological evaluation. Membranes harvested from transiently transfected Cos-7 cells exhibited high affinity, saturable [¹²⁵I]PYY binding. The time course of specific binding was measured in the presence of 0.06 nM ¹²⁵I-PYY 35 (Fig. 5). The association curve was monophasic, with an observed association rate (K_{obs}) of $0.12 \pm 0.02 \text{ min}^{-1}$ and a $t_{1/2}$ of 6 min; equilibrium binding was 95% complete

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within 26 min and 100% complete within 50 min (n = 3). For comparison, we also measured the time course of binding to human Y1 receptors transiently expressed in COS-7 cells. The association curve was monophasic, with 5 a K_{obs} of $0.06 \pm 0.02 \text{ min}^{-1}$ and a $t_{1/2}$ of 12 min; equilibrium binding was 95% complete within 51 min and 100% complete within 90 min (n = 3) (data not shown). The different patterns of radioligand association for hp25a and human Y1 receptors suggest novel mechanisms of receptor/ligand 10 interaction.

Saturation binding data for ^{125}I -PYY were fit to a one-site model with an apparent K_d of $0.11 \pm 0.01 \text{ nM}$ and an apparent B_{max} of $1.42 \pm 0.05 \text{ pmol/mg membrane protein}$, 15 corresponding to approximately 1.4×10^5 receptors/cell (n = 4; Fig. 6). Given that the transfection efficiency was 20-30% (data not shown), the receptor density on transfected cells was probably closer to 7×10^5 /cell. Membranes from mock-transfected cells, when prepared and 20 analyzed in the same way as those from hp25a-transfected cells, displayed no specific binding of ^{125}I -PYY. We conclude that the ^{125}I -PYY binding sites observed under the described conditions were derived from the hp25a construct.

25 The pharmacological profile of hp25a was defined by membrane binding assays. The receptor was probed for features of all well characterized pancreatic polypeptide family receptors including Y1, Y2, Y3, and PP. The rank 30 order of affinity for several peptide analogs was derived from competitive displacement of ^{125}I -PYY (Fig. 7 and Table 2). The hp25a receptor was compared with two model systems: 1) the cloned human Y1 receptor (Larhammar et al., 1992; Herzog et al., 1992) transiently expressed in 35 COS-7 cells, and 2) the Y2-like receptor population expressed by human SK-N-Be(2) neuroblastoma cells (Wahlestedt et al., 1991; Dumont et al., 1992). No

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models for human Y3 and human PP receptors have been described.

PP bound to hp25a with extremely high affinity ($K_i = 0.029$ nM) and dramatic selectivity: PP was > 6000 -fold selective for hp25a over human Y1 receptors ($K_i = 200$ nM) and SK-N-Be(2) receptors ($K_i > 300$ nM). This profile suggests that hp25a could function selectively as a PP receptor *in vivo*. The data further indicated, however, that hp25a bound quite well to human NPY ($K_i = 1.4$ nM) and even better to human PYY ($K_i = 0.62$ nM). These K_i values, while lower than the K_i for PP, are comparable to the effective concentrations of NPY and PYY from numerous physiological and pharmacological studies (Dumont, 1992). In our investigation, SK-N-Be(2) receptors bound human NPY and human PYY in the same rank order as hp25a but with 5- to 10-fold higher affinity, whereas human Y1 receptors bound human NPY and human PYY in the opposite rank order with 5- to 30-fold higher affinity. Hydrolysis of the carboxy terminal amide to free carboxylic acid, as in human NPY free acid, was disruptive for binding to all receptors. A requirement for a carboxy terminal amide appears to be a common structural feature of all pancreatic polypeptide family peptide/receptor interactions.

Fuhlendorff and co-workers replaced Ile³¹ and Gln³⁴ in NPY with the corresponding residues from PP to create [Leu³¹, Pro³⁴]NPY, which is commonly used to distinguish Y1 from Y2 receptors (Fuhlendorff, 1990). Human [Leu³¹, Pro³⁴]NPY displayed > 2300 -fold selectivity for human Y1 receptors over SK-N-Be(2), but only 5-fold selectivity for human Y1 receptors over hp25a. Human [Leu³¹, Pro³⁴]NPY was a better ligand for hp25a ($K_i = 0.60$ nM) than was human NPY itself ($K_i = 1.4$ nM). This is possibly a reflection of the way in which [Leu³¹, Pro³⁴]NPY mimics PP at positions 31 and 34. In contrast, the

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[Leu³¹, Pro³⁴]NPY analog was well tolerated by the human Y1 receptor ($K_i = 0.13$ nM), but not preferred over the parent peptide ($K_i = 0.049$ nM).

5 hp25a displayed an intermediate level of sensitivity to N-terminal deletions of NPY and PYY, less so than human Y1 receptors. Removing Tyr¹ from porcine NPY resulted in a 29-fold loss in affinity for human Y1 receptors when compared with the full length parent peptide. The same
10 modification decreased affinity 4-fold for hp25a receptors and 3-fold for SK-N-Be(2) receptors. It is interesting in this regard that human PP contains Ala¹; the Tyr¹ of NPY may not play much of a role in receptor recognition. Truncation to NPY₁₃₋₃₆ decreased affinity
15 1000-fold for human Y1 receptors, 33-fold for hp25a, and 4-fold for SK-N-Be(2) receptors. Further truncation to porcine NPY₂₂₋₃₆ decreased affinity 3500-fold for human Y1 receptors, 120-fold for hp25a, and 11-fold for SK-N-Be(2) receptors. In this regard, the hp25a receptor
20 shares features of both Y1- and Y2-like pharmacology, as would be expected if the N-terminal region of porcine NPY were only moderately involved in receptor recognition.

An important structural difference between human PP,
25 human PYY and human NPY is that both human NPY and PYY contain Gln³⁴, whereas human PP contains Pro³⁴. When Gln³⁴ in NPY was replaced with Pro³⁴ (as in the analog [Leu³¹, Pro³⁴]NPY), an increase in binding affinity for the human Y4 receptor was observed. A similar increase in
30 binding affinity was detected when Gln³⁴ of PYY was replaced with Pro³⁴, supporting the proposal that PP-like peptides are preferred by the Y4 receptor. Replacement of Pro³⁴ in human PP by Gln³⁴ (as in [Ile³¹, Gln³⁴]PP) caused very little change in PP binding affinity,
35 however, suggesting that in the case of PP there are significant contributions to binding affinity from other regions of the peptide structure.

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Applicant's further extended the structure/activity data for human PP fragments (PP₂₋₃₆, PP₁₃₋₃₆, PP₂₀₋₃₆, PP₂₇₋₃₆, and PP₃₁₋₃₆). PP binding was unaffected by N-terminal truncation to PP₂₋₃₆, but further truncation to PP₁₃₋₃₆ and 5 beyond was disruptive. The shortest PP fragment tested, PP₃₁₋₃₆, bound selectively to the Y4 receptor with $K_i = 350$ nM, and hydrolysis of the C-terminal amide was detrimental ($K_i > 10,000$ nM for human PP₃₁₋₃₆ free acid), as reported earlier for NPY. We conclude that the binding 10 of PP to the Y4 receptor resembles the binding of NPY to the Y1 receptor, in that 1) Pro³⁴ is well-tolerated and 2) both ends of the peptide are required for optimal binding activity. This is in contrast to the Y2 binding model, in which 1) Pro³⁴ is not well-tolerated and 2) the 15 N-terminal region of NPY does not contribute significantly to binding affinity. Note also that the Y2-selective ligands human PYY₃₋₃₆ and C2-NPY display relatively low affinity for the human Y4 receptor.

20 Additionally, the binding of the tetrapeptide invertebrate neurotransmitter Phe-Met-Arg-Phe-Amide (FMRF-amide) was investigated. This peptide has been shown to mimic several functions of NPY including the stimulation of food intake in rats (Robert, 1988). 25 FMRFamide bound selectively to the Y4 receptor with a K_i value of 4000 nM. A closely related derivative, Phe-Leu-Arg-Phe-amide (FLRFamide), displayed improved Y4 binding affinity ($K_i = 750$ nM) while maintaining selectivity. We also investigated the binding of [D-Trp³²]NPY. This 30 peptide was reported to stimulate food intake when injected into rat hypothalamus, and also to attenuate NPY-induced feeding in the same paradigm (Balasubramaniam, 1994). [D-Trp³²]NPY displayed relatively low binding affinity for the human Y4 receptor 35 as well as for the human Y1 and Y2 receptor subtypes. Data for these and other new peptides not included in the original patent filing are listed in Table 3.

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Untransfected NIH-3T3 and LM(tk-) were pre-screened for specific ^{125}I -PYY binding and found to be negative (data not shown). After co-transfection with the human Y4 cDNA and a G418-resistant gene and selection with G-418, 5 surviving colonies were screened for specific binding of ^{125}I -PYY. Two positive clones were identified and isolated for further study (NIH-3T3 hY4 clone #5 and LM(tk-) hY4 clone #3). The binding of ^{125}I -PYY to membranes from the NIH-3T3 stable clone was saturable over a radioligand 10 concentration range of 0.5 pM to 2.5 nM. Binding data were fit to a one-site binding model with an apparent K_d of 0.17 nM \pm 0.005 and a receptor density of 350 \pm 80 fmol/mg membrane protein (mean \pm s.e.m., n = 2). The LM(tk-) clone displayed an estimated receptor density of 15 7 fmol/mg membrane protein during the primary selection screen and was not analyzed further in a saturation assay.

Activation of all Y-type receptors described thus far is 20 thought to involve coupling to pertussis toxin-sensitive G-proteins which are inhibitory for adenylate cyclase activity (G_i or G_o) (Wahlestedt and Reis, 1993). Based on these prior observations, we investigated the ability of PP to inhibit forskolin-stimulated cAMP accumulation in 25 LM(tk-) cells stably expressing the human Y4 receptor. Incubation of intact cells with 10 μM forskolin produced -10-fold increase in cAMP accumulation over a 5 minute period, as determined by radioimmunoassay. Simultaneous incubation with human PP decreased the forskolin- 30 stimulated cAMP accumulation by 67% in stably transfected LM(tk-) cells (Fig. 8) but not in untransfected cells (data not shown). Applicants conclude that human Y4 receptor activation can result in decreased cAMP accumulation, very likely through inhibition of adenylate 35 cyclase activity.

Peptides selected for their ability to bind to the

transiently expressed human Y4 receptor were investigated for their ability to activate the human Y4 in the cAMP assay (Table 4). Note that both human PP and human PP₂₇₋₃₆ bound the Y4 receptor with a K_i value of 0.06 nM, and that 5 each displayed comparable activity in the cAMP assay with closely matching EC₅₀ values of 0.09 nM and 0.08 nM, respectively. The truncated PP fragments PP₂₇₋₃₆ and PP₃₁₋₃₆ were relatively weak ligands in the binding assay and were also less than 50% as effective as the full length 10 PP in reducing forskolin-stimulated cAMP, thereby acting as partial agonists. Similarly, both NPY and PYY (which deviate from PP primarily in the N-terminal- regions) yielded EC₅₀ values \geq 10-fold larger than their K_i values. Receptor activation (more so than binding) may 15 therefore depend heavily upon N-terminal PP structure. The functional activity of the reported feeding behavior modulator [D-Trp³²]NPY was also investigated. Consistent with this peptide's low binding affinity for the human Y4 receptor, no functional activity of the peptide was 20 detected at concentrations up to 0.3 uM (see Table 4), or when tested at 0.3 uM for antagonism of the PP functional response (data not shown).

The intracellular free calcium concentration was markedly 25 increased in LM(tk-) cells stably transfected with the human Y4 receptor after application of 100 nM human PP (Δ [Ca²⁺]_i = 325 nM; Fig. 9). The response to 100 nM NPY was relatively small (Δ [Ca²⁺]_i = 68 nM). Untransfected LM(tk-) cells were unresponsive to either peptide (data 30 not shown). When human PP was further analyzed in a concentration/response curve, the maximum Δ [Ca²⁺]_i measured was 334 nM and the EC₅₀ was 35 nM (Fig. 9, Inset). This greater activity of PP over NPY is consistent with the pharmacological profiles derived 35 from both binding and cAMP assays described above. The calcium mobilization assay thereby provides a second pathway through which Y4 receptor activation can be

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measured.

Y4 mRNA was detected by PCR techniques in a broad range of human tissues. Relatively intense hybridization signals were detected in total brain, coronary artery, and ileum, suggesting a potential role for Y4 receptors in CNS function, cardiovascular regulation, and gastrointestinal physiology (Table 5).

10 The cDNA corresponding to the rat Y4 homolog was transiently expressed in COS-7 cells for membrane binding studies. The binding of ^{125}I -PYY to the rat Y4 receptor was saturable over a radioligand concentration of 0.5 pM to 2.5 nM. Binding data were fit to a one-site model with an apparent K_d of $0.15 \text{ nM} \pm 0.005$ and a receptor density of $275 \pm 3 \text{ fmol/mg membrane protein}$ (mean \pm s.e.m., n = 2). As determined by using peptide analogs within the pancreatic polypeptide family, the rat Y4 pharmacological profile bears a resemblance to the human Y4 receptor; 15 there are several interesting exceptions, however, including frog PP, salmon PP, human PP₃₁₋₃₆, and avian PP, each of which discriminated ~10-fold between the rat and human receptor subtypes (Table 6). The differences may reflect the fact that PP is not well conserved among 20 species relative to NPY and PYY; hence the species homologs of PP are likely to exhibit more variability in ligand binding.

25 In summary, both the human Y4 receptor and the rat Y4 receptor displayed features unique among the neuropeptide receptors, exhibiting a profile which is divergent from their closest relatives, Y1 or Y2, in that each binds optimally to PP rather than to NPY or PYY (see Tables 1, 2 and 6). Unlike the Y1 and Y2 receptor models, the Y4 receptor appears to be a reasonable target for all three peptide ligands.

TABLE I

Pharmacologically defined receptors for NPY and related pancreatic polypeptides.

Rank orders of affinity are based on published reports of binding and functional data (Wahlestedt et al., 1990; Schwartz et al., 1991; Wahlestedt et al., 1993; Dumont et al., 1992). Missing peptides in the series reflect a lack of published information.

Receptor	Affinity (-pK _i or -pEC ₅₀)					
	11 to 10	10 to 9	9 to 8	8 to 7	7 to 6	< 6
Y1	NPY PYY [Leu ¹¹ , Pro ¹⁴]N PY	NPY ₂₋₃₆	NPY ₁₃₋₃₆	PP		
Y2		PYY NPY NPY ₂₋₃₆	NPY ₁₃₋₃₆		[Leu ¹¹ , Pro ¹⁴]N PY PP	
Y3		NPY	[Pro ¹⁴]NPY	NPY ₁₃₋₃₆ PP		PYY
PP	PP		[Leu ¹¹ , Pro ¹⁴]N PY		NPY	

TABLE 2
Pharmacological profile of the hp25a receptor.

Binding data reflect competitive displacement of ^{125}I -PYY from membranes of COS-7 cells transiently expressing hp25a receptors. Peptides were tested at concentrations ranging from 0.001 nM to 100 nM. IC_{50} values corresponding to 50% displacement were determined by nonlinear regression analysis and converted to K_i values according to the equation, $K_i = \text{IC}_{50}/(1 + [L]/K_d)$, where $[L]$ is the ^{125}I -PYY concentration and K_d is the equilibrium dissociation constant of ^{125}I -PYY. The data shown are representative of at least two independent experiments.

Competitor	Human Y1, K_i (nM)	hp25a, K_i (nM)	SK-N-Be (2), K_i (nM)
human PP	200 \pm 68	0.029 \pm 0.006	> 300
human $[\text{Leu}^{11, \text{Pro}34}]$ NPY	0.13 \pm 0.02	0.60 \pm 0.09	> 300
human PPY-PYY	0.085 \pm 0.021	0.62 \pm 0.15	0.11 \pm 0.02
porcine NPY	0.049 \pm 0.001	1.2 \pm 0.2	0.28 \pm 0.04
human NPY	0.049 \pm 0.009	1.4 \pm 0.1	0.13 \pm 0.02
porcine NPY ₂₋₃₆	1.4 \pm 0.2	4.4 \pm 1.3	0.41 \pm 0.09
porcine NPY ₁₃₋₃₆	51 \pm 16	39 \pm 5	1.8 \pm 0.4
porcine PY ₁₃₋₃₆	32 \pm 7	47 \pm 6	0.86 \pm 0.14
porcine NPY ₁₆₋₃₆	45 \pm 4	54 \pm 2	5.0 \pm 0.5
porcine NPY ₁₈₋₃₆	28 \pm 5	63 \pm 7	2.1 \pm 0.5
human NPY free acid	> 300	79 \pm 17	280 \pm 120
porcine NPY ₂₀₋₃₆	62 \pm 6	100 \pm 20	3.1 \pm 0.6
porcine NPY ₂₂₋₃₆	170 \pm 30	140 \pm 63	3.2 \pm 0.6
porcine NPY ₂₆₋₃₆	> 300	> 300	70 \pm 7

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Table 3: human Y4 receptor vs. Y-type receptors cloned from human.

Binding data reflect competitive displacement of ^{125}I -PYY from membranes of COS-7 cells transiently expressing human Y1, human Y2, and human Y4 receptors. IC_{50} values corresponding to 50% displacement were determined by nonlinear regression analysis and converted to K_i values according to the equation Chang-Prusoff equation, $K_i = \text{IC}_{50}/(1 + [L]/K_d)$, where [L] is the ^{125}I -PYY concentration and K_d is the equilibrium dissociation constant of ^{125}I -PYY. Any peptide not included in the original patent filing is referred to as a "new peptide".

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Table 3

Peptide	Y1	Y2	Y4	Comments
PP, human	77	> 1000	0.06	
PP ₂₋₃₆ , human	> 40	> 100	0.06	new peptide
PP ₁₃₋₃₆ , human	> 100	> 100	39	new peptide
PP ₂₀₋₃₆ , human	> 100	> 100	> 100	new peptide
PP ₂₇₋₃₆ , human	> 100	> 100	> 88	new peptide
PP ₃₁₋₃₆ , human	> 10000	> 10000	350	new peptide
PP ₃₁₋₃₆ free acid, human	> 10000	> 10000	> 10000	new peptide
Phe-Met-Arg- Phe-Amide	12000	75000	4000	
Phe-Leu-Arg- Phe-Amide	15000	> 10000 0	750	new peptide
[Ile ³¹ , Gln ³⁴]PP, human	> 86	20	0.09	new peptide
PP, bovine	240	> 820	0.05	new peptide
PP, rat	460	> 1000	0.18	new peptide

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Table 3 continued

Peptide	Y1	Y2	Y4	Comments
PP, salmon	0.20	0.17	3.2	new peptide
PP, avian	400	>	7.0	new peptide
		1000		
PP, frog	98	>	61	new peptide
		1000		
PYY, human	0.19	0.36	0.87	
PYY, porcine	0.14	0.35	1.3	new peptide
PYY ₃₋₃₆ , human	45	0.70	14	new peptide
PYY ₁₃₋₃₆ , porcine	33	1.5	46	
[Pro ³⁴]PYY, human	0.14	> 310	0.12	new peptide
Peptide	Y1	Y2	Y4	Notes
NPY, human	0.08	0.74	2.2	
NPY, porcine	0.07	0.81	1.1	
Melanostatin (frog NPY)	0.07	0.87	1.2	new peptide
NPY ₂₋₃₆ , human	3.6	2.0	16	new peptide
NPY ₂₋₃₆ , porcine	2.4	1.2	5.6	
NPY ₁₃₋₃₆ , porcine	70	2.5	38	

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Table 3 Continued

Peptide	Y1	Y2	Y4	Comments
NPY ₁₆₋₃₆ , porcine	41	3.6	54	
NPY ₁₈₋₃₆ , porcine	70	4.2	> 290	
NPY ₂₀₋₃₆ , porcine	63	3.6	120	
NPY ₂₂₋₃₆ , porcine	> 1000	18	> 990	
NPY ₂₆₋₃₆ , porcine	> 1000	380	304	
[Leu ³¹ , Pro ³⁴]NPY, human	0.15	> 120	1.1	
[Leu ³¹ , Pro ³⁴]NPY, porcine	0.15	> 540	1.5	new peptide
O-Me-Tyr ²¹ - NPY, human	0.12	1.55	6.1	new peptide
NPY free acid, human	490	>	> 1000	
		1000		
NPY ₁₋₂₄ amide, human	> 1000	>	> 1000	new peptide
		1000		
C2-NPY, porcine	73	3.5	120	new peptide
[D-Trp ³²]NPY, human	> 1000	>	> 1000	new peptide
		1000		

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TABLE 4: Functional activation of the human Y4 receptor and inhibition of cAMP accumulation.

K_i values were derived from binding assays as described in Table 3. Peptides were evaluated for binding affinity and then analyzed for functional activity. Functional data were derived from radioimmunoassay of cAMP accumulation in stably transfected LM(tk-) cells stimulated with 10 μ M forskolin. The maximum inhibition of cAMP accumulation relative to that produced by human PP (E_{max}) and the concentration producing a half-maximal effect (EC_{50}) were determined by nonlinear regression.

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Table 4

Peptide	Binding		Function
	K _i (nM)	EC ₅₀ (nM)	
PP, human	0.06	0.09	100 %
PP ₂₋₃₆ , human	0.06	0.08	101%
PP ₁₃₋₃₆ , human	39	580	96%
PP ₂₇₋₃₆ , human	> 88	3500	50 %
PP ₃₁₋₃₆ , human	> 10000	89000	47 %
[Ile ³¹ ,Gln ³⁴]PP , human	0.09	0.27	101%
salmon PP	3.2	110	96%
PYY, human	0.87	47	118%
[Pro ³⁴]PYY, human	0.12	1.1	106%
NPY, human	2.2	20	98%
NPY, porcine	1.1	68	105%
NPY ₁₈₋₃₆	> 290	Not detected	
[Leu ³¹ ,Pro ³⁴]N PY, human	1.1	35	105%
[Leu ³¹ ,Pro ³⁴]N PY, porcine	1.5	26	111%

Table 4 continued

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Peptide	Binding	Function
[D- Trp ³²]NPY, human	> 1000	Not detected

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TABLE 5: Macrolocalization of Y4 receptor mRNA in human tissues by PCR.

Localization data reflect PCR-based amplification of human Y4 cDNA derived from mRNA extracts of human tissues. Southern blots of the PCR products were prepared and hybridized with ^{32}P -labeled oligonucleotide probes selective for Y-type receptor subtypes. The labeled products were recorded on X-ray film and the relative signal density was determined by visual inspection. In this rating scheme, + = faint signal, + + = moderate signal, + + + = intense signal.

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Table 5

Human tissues	human Y4 PCR product
total brain	+++
frontal brain	+
ventricle (heart)	++
atrium (heart)	+
thoracic aorta	++
coronary artery	+++
nasal mucosa	+
mesentery	++
stomach	++
ileum	+++
pancreas	not determined
liver	(-)
kidney	not determined
bladder	+
penis	+
testes	+
uterus (endometrium)	++
uterus (myometrium)	+

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TABLE 6: Pharmacological binding profile of the rat Y4 receptor vs. the human Y4 receptor.

Binding data reflect competitive displacement of ^{125}I -PYY from membranes of COS-7 cells transiently expressing rat Y4 and human Y4 receptors. IC₅₀ values corresponding to 50% displacement were determined by nonlinear regression analysis and converted to K_i values according to the equation Chang-Prusoff equation, K_i = IC₅₀ / (1 + [L] / K_d), where [L] is the ^{125}I -PYY concentration and K_d is the equilibrium dissociation constant of ^{125}I -PYY.

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Table 6

Peptide	Rat Y4	Human Y4
PP, human	0.12	0.06
PP, rat	0.20	0.18
PP, bovine	0.15	0.05
PP, frog	0.19	62
PP, salmon	0.36	3.2
PP ₃₁₋₃₆ , human	20	350
PP, avian	> 82	7
PP ₃₁₋₃₆ free acid, human	> 100	> 10000
PYY, porcine	0.58	1.3
NPY, human	1.7	2.2
NPY, porcine	1.8	1.1
NPY ₂₋₃₆ , human	5	16
NPY ₁₃₋₃₆ , porcine	135	38
[Leu ³¹ ,Pro ³⁴]NPY, human	0.59	1.2
NPY free acid, human	> 1000	> 1000
C2-NPY, porcine	22	120
[D-Trp32]NPY, human	> 1000	> 1000

Discussion

Applicants have cloned DNA representing a novel human neuropeptide Y/peptide YY/pancreatic polypeptide receptor (Y4) from human genomic DNA. Of all known G protein-coupled receptor sequences (EMBL/Genbank Data Base), the greatest homology was displayed between hp25a and the Y1 receptor genes (mouse--Eva et al., 1992; rat--Eva et al., 1990; and human--Larhammar et al., 1992). Comparison of the human hp25a deduced amino acid sequence with known G protein-coupled receptor sequences indicates the greatest concentration of identical amino acids to be in the transmembrane domains. In these TM regions, the percentage of identity for hp25a clone is 55% compared to human Y1, and less than 35% with other members of the peptide subfamily and other G protein-coupled receptor subfamilies. The alignment of this human hp25a sequence, relative to other G protein-coupled receptors or other members of the neuropeptide receptor subfamily, specifically human Y1, indicates a unique sequence, proving hp25a is a newly characterized receptor. The homology of hp25a to Y1 indicates that it is related to the NPY/PYY/PP family of receptors.

While the hp25a human receptor sequence exhibits higher overall and transmembrane identity to the rs16b rat Y4 receptor sequence than to other Y-type receptors such as the human Y1 receptor, the divergence between the rat Y4 and human Y4 sequences may contribute to the pharmacological differences between the two receptors. The isolation of the rat homologue of the Y4 receptor provides the means to compare the pharmacological properties of the rat and human Y4 receptors (see below) in relation to their observed differences in primary structures. These data will be critical to the design and testing of human therapeutic agents acting

at these sites.

The unique pharmacological profile of the hp25a human Y4 receptor suggests that this receptor can serve as a 5 novel target for the development of subtype selective ligands. The competitive displacement studies indicate that human PP is the preferred ligand for hp25a. The receptor also binds with high affinity to human NPY and human PYY, which share \geq 47% amino acid identity with 10 human PP. Affinity is enhanced by modifying NPY to closely resemble PP, as in [Leu³¹, Pro³⁴]NPY. Decreased affinity for C-terminal fragments of NPY suggest that both N- and C-terminal regions of NPY contribute to hp25a receptor recognition. hp25a was less sensitive to 15 N-terminal deletion of NPY than was the human Y1 receptor. One may speculate that both Y1 and hp25a share a common mechanism of peptide interaction which has been optimized for either NPY or PP, respectively.

20 The pharmacological data do not support classification of hp25a as a Y1 receptor, in which case it would display $>$ 4000-fold selectivity for binding to human NPY over human PP (Table 2). Neither do the data support classification as a Y2 receptor, in which case 25 it would tolerate N-terminal deletion of NPY but not exchange of Gln³⁴ for Pro³⁴ (Table 2). Finally, the data fails to support the classification of hp25a as a Y3 receptor, since it would be expected to display greater affinity for NPY than for PP or PYY (Wahlestedt et al., 30 1991). Therefore, applicants are designating the hp25a receptor as a Y4 receptor.

The additional data included here reflect an increased 35 understanding of receptor ligand/interactions. Our further characterization of Y4 receptor pharmacology has indicated, for example, that the binding affinity

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for either human NPY ($K_i = 2.2$ nM) or human PYY ($K_i = 0.87$ nM) can be enhanced by conversion to human [^{Leu⁹} Pro³⁴]NPY ($K_i = 1.1$ nM) or human [Pro³⁴]PYY ($K_i = 0.14$ nM). This information supports the importance of
5 Pro³⁴- in the peptide pharmacophore and could potentially be incorporated into the design of metabolically stable nonpeptide ligands with Y4-selectivity. Additionally, the data prompt a re-evaluation of literature reports in which [Pro³⁴]PYY is
10 described as a Y1-selective ligand. Our results indicate that [Pro³⁴]PYY does not discriminate between the cloned human Y1 and cloned human Y4 receptor ($K_i = 0.12$ and 0.14 nM, respectively) such that [Pro³⁴]PYY cannot be used in isolation to define receptor
15 subtypes.

Other particularly interesting peptides include FMRF-amide, FLRF-amide, and [D-Trp³²]NPY. FMRF-amide and [D-Trp³²]NPY have both been shown to modulate food intake
20 in rats (get ref from George M.). While FMRF-amide and its derivative displayed some degree of Y4-selectivity (albeit relatively low affinity compared to human PP), [D-Trp³²]NPY was essentially inactive at all Y-receptor subtypes studied. These profiles must be
25 considered as efforts are undertaken to validate the receptor mechanism of NPY-induced food intake. The tetrapeptide FLRF-amide has additional value as a starting point for the design of small nonpeptide compounds with Y4 selectivity.

30 Applicants now have several Y4 receptor expression systems from which to chose, each uniquely suited to different research questions. The transient expression system in COS-7, for example, allows one to generate
35 sufficient quantities of membranes for routine structure/activity relationship questions. Applicants can also produce mutant receptors by site-directed

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mutagenesis or other mutagenesis techniques and express them transiently in COS-7 for a comparison of pharmacological properties with those of the wild-type receptor. In this way, one can gain insight into 5 receptor binding pockets, ligand binding domains, and mechanisms of activation. Whereas the transient expression system requires a new transfection for every cell or membrane harvest, the stable expression system offers the convenience of a single transfection step 10 followed by routine passaging techniques. The stable system also offers the opportunity to select receptor density, which could be an important factor in evaluating the intrinsic activity of Y4 receptor ligands.

15 Applicants' characterization of the stably expressed Y4 receptor now shows definitively that the Y4 receptor can couple simultaneously to both cAMP regulation and calcium mobilization in a single cell type. The EC₅₀ 20 for the calcium response is significantly higher than the EC₅₀ for the cAMP response, suggesting that calcium mobilization may reflect promiscuous coupling of the receptor to G-protein other than that required for cyclase regulation. The functional assays allow one to 25 assign agonist and antagonist activities to receptor selective compounds and thereby provide one with critical tools for drug design.

30 The question logically arises as to whether hp25a should be classified as a PP receptor. To applicants' knowledge, no human PP receptor has been described. One must therefore look to the rat PP receptor for comparison. The rat PP receptor bound PP and analogs in the same rank order as hp25a (PP > [Leu³¹, Pro³⁴]NPY > 35 NPY) (Schwartz et al., 1990). The rat PP receptor also appeared to bind both N- and C-terminal regions of the peptide ligand (Schwartz et al., 1987). A glaring

discrepancy between hp25a and the rat PP receptor is that the latter displayed > 10,000-fold selectivity for PP over NPY (Schwartz et al., 1990).

In applicants' localization experiments Y4 mRNA was 5 detected by PCR techniques in a broad range of human tissues. Relatively intense hybridization signals were detected in total brain, coronary artery, and ileum, suggesting a potential role for Y4 receptors in CNS function, cardiovascular regulation, and 10 gastrointestinal physiology. This localization pattern is consistent with previously reported studies of PP-mediated effects at 1) brainstem sites (McTigue et al., 1993; Whitcomb et al., 1990), 2) on arterial blood pressure (Wager-Page et al., 1993a) and 3) on gastric 15 acid secretion and gastrointestinal motility (McTigue et al., 1993; Wager-Page et al., 1993b). A more definitive localization of the Y4 receptor mRNA and receptor expression (i.e., whether on enterocytes, vascular smooth muscle cells, neurons, etc.) is 20 attainable through *in situ* hybridization and receptor autoradiography techniques. There are to applicants' knowledge no published reports of PP receptor localization in human tissue as obtained through binding or functional studies. It may be informative, 25 however, to compare the human Y4 macrolocalization data presented here with PP receptor characterization in the rat. PP receptors have been described, for example, in brainstem nuclei such as the area postrema, interpeduncular nucleus, dorsomedial nucleus, and the 30 nucleus tractus solitarius (Whitcomb et al., 1990), consistent with the identification of Y4 mRNA in human brain. The PP receptors in rat brain stem are accessible to circulating PP, which is released upon vagal stimulation of the pancreas during feeding 35 (Whitcomb et al., 1990). Activation of brainstem PP receptors inhibits further pancreatic secretion, increases gastric acid secretion, enhances gastric

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motility, and increases gastric emptying time (Louie et al., 1985; McTigue and Rogers, 1993). A Y4 receptor antagonist then, would be expected to slow down gastric emptying time and potentially reduce meal size.

5

Given the similarities in pharmacologic profiles between the published PP receptor and the hp25a human Y4 receptor, it would be tempting to call hp25a the human PP receptor. Applicants believe that calling 10 hp25a the human PP receptor, however, would be misleading. This is because the relatively compressed window of affinity for PP, PYY, and NPY ($0.02 \text{ nM} \leq K_i \leq 1.5 \text{ nM}$) makes hp25a a potential target for all three peptide ligands. Future localization experiments may 15 help resolve the relationship between hp25a and the PP receptor.

Applicants propose that hp25a be known as the Y4 receptor. The name is not biased toward any one member 20 of the pancreatic polypeptide family. The "Y" has its roots in the original classification of Y1 and Y2 receptor subtypes (Wahlestedt et al., 1987). The letter reflects the conservation in pancreatic polypeptide family members of the C-terminal tyrosine, 25 described as "Y" in the single letter amino acid code. Applicants note that the cloned human Y1 receptor was introduced by Larhammar and co-workers as a "human neuropeptide Y/peptide YY receptor of the Y1 type", with peptide ligands listed in rank order of affinity 30 (Larhammar et al., 1992). Similarly, hp25a could be described as a human pancreatic polypeptide/peptide YY/neuropeptide Y receptor of the Y4 type.

hp25a is to applicants' knowledge the first "Y type" 35 receptor to be cloned from a subtype family other than Y1. The reported Y3 receptor cloned from bovine brain (Rimland et al., 1991) was later described as having

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been misidentified (Jazin et al., 1993; Herzog et al., 1993). A Y2-like receptor (PR4) was cloned from drosophila and characterized using mammalian analogs of NPY (Li et al., 1992); however, the classification of 5 this receptor is controversial. The receptor was relatively insensitive to NPY; concentrations ranging from 0.3 to 10 μ M were required to elicit calcium mobilization in oocytes injected with PR4 mRNA (Li et al., 1992). The receptor also displayed a rank order 10 of potency for NPY analogs distinct from that observed in mammalian systems (Wahlestedt et al., 1993; Li et al., 1992). Furthermore, an NPY analog has not been isolated from drosophila (Wahlestedt et al., 1993). It is possible that an unidentified ligand in drosophila 15 can activate PR4 more readily than NPY, and as such, the receptor may eventually be reclassified.

The cloning and expression of a Y4 (hp25a) receptor represents a major advance in the ability to analyze 20 numerous physiological processes mediated by the pancreatic polypeptide family. Binding sites for PP, PYY, or NPY have a widespread anatomical distribution in peripheral targets such as neuromuscular junction, smooth muscle, stomach chief cells, intestinal 25 enterocytes, kidney proximal tubule, and fat cells (Dumont et al., 1992; Castan et al., 1992). These receptors are therefore in a position to potentially regulate a variety of physiological functions including cognition, circadian rhythm, EEG synchronization, body 30 temperature, blood pressure, locomotor activity, neuroendocrine release, sexual/reproductive behavior, feeding, sympathetic activation, sensory transmission, gastrointestinal function, intestinal secretion, renal absorption, and cardiovascular function (Wahlestedt et 35 al., 1993).

Y4 receptors are an invaluable resource for drug

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design. The pancreatic polypeptide family is potentially involved in several pathophysiological conditions including memory loss, depression, anxiety, epileptic seizure, pain, hypertension, locomotor problems, circadian rhythm disorders, eating/body weight disorders, sexual/reproductive disorders, nasal congestion, and diarrhea (Wahlestedt et al., 1993; Dumont et al., 1992). The available data implicate this receptor in the control of obesity and other disorders of feeding including bulimia and anorexia. The chemical synthesis of selective drugs not only for Y4 but for all "Y type" receptors will be greatly accelerated by preliminary screening against a homogeneous population of cloned human Y4 receptors. As more specific pharmacological tools become available for probing receptor function, additional therapeutic indications are likely to be discovered.

Applicants do not know whether hp25a represents the single Y4 receptor expressed in the human genome, or whether there exists a group of structurally related Y4 receptor subtypes. This is an issue which can be resolved using nucleotide sequences from Y4 receptor as the basis for *in situ* localization, antisense or "knockout" strategies, homology cloning, and related techniques. Such approaches will enable one to investigate the existence of potentially novel receptor subtypes with pharmacologic and therapeutic significance.

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In conclusion, the primary structure of the proteins encoded by hp25a (Y4) gene and its homolog in the rat, as well as its unique pharmacological profile obtained for the Y4 receptor subtype, indicate that these genes represent a new pancreatic polypeptide receptor subfamily. Additional cloning efforts will be required to isolate additional members of this newly recognized

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neuropeptide receptor family.

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: Bard, Jonathan A.
 Walker, Mary
 Branchek, Theresa
 Weinshank, Richard L.

(ii) TITLE OF INVENTION: DNA ENCODING A HUMAN NEUROPEPTIDE Y/PEPTIDE YY/PANCREATIC POLYPEPTIDE RECEPTOR (Y4) AND USES THEREOF

(iii) NUMBER OF SEQUENCES: 28

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(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
 (B) COMPUTER: IBM PC compatible
 (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 (D) SOFTWARE: PatentIn Release #1.0, Version #1.25

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:
 (B) FILING DATE:
 (C) CLASSIFICATION:

(viii) ATTORNEY/AGENT INFORMATION:

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 (B) REGISTRATION NUMBER: 28,678
 (C) REFERENCE/DOCKET NUMBER: 44743-A-PCT\JPW\MAT

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 (B) TELEFAX: (212) 391-0525

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1320 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION: 88..1212

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60

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111

Met Asn Thr Ser His Leu Leu Ala

- 95 -

1 5

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Gly Thr Pro Tyr Asn Phe Ser Glu His Cys Gln Asp Ser Val Asp Val	
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GAC TAC TGG ATC TTT GGA GAG ACC CTC TGC AAG ATG TCG GCC TTC ATC	447
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Val Leu Ser Leu Pro Phe Leu Ala Asn Ser Ile Leu Glu Asn Val Phe	
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His Lys Asn His Ser Lys Ala Leu Glu Phe Leu Ala Asp Lys Val Val	
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Cys Thr Glu Ser Trp Pro Leu Ala His His Arg Thr Ile Tyr Thr Thr	
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Val Val Leu Val Val Met Val Val Ala Phe Ala Val Leu Trp Leu Pro	
265 270 275 280	

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(ii) MOLECULE TYPE: protein	
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-97-

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 260 265 270
 Ala Phe Ala Val Leu Trp Leu Pro Leu His Val Phe Asn Ser Leu Glu
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 Asp Trp His His Glu Ala Ile Pro Ile Cys His Gly Asn Leu Ile Phe
 290 295 300
 Leu Val Cys His Leu Leu Ala Met Ala Ser Thr Cys Val Asn Pro Phe
 305 310 315 320
 Ile Tyr Gly Phe Leu Asn Thr Asn Phe Lys Lys Glu Ile Lys Ala Leu
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(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 45 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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45

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 45 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

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(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

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(2) INFORMATION FOR SEQ ID NO:5:

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- (A) LENGTH: 45 base pairs
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- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

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45

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- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

CAGCAAGTCT GAGAAGGAGA GGTTCACCGAT CAGAATGTTG GTGAC

45

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 54 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

TGCAGAACTGA ATCCTTTGT GCAATGCGTC TCCATTACAG TATCCATTCT

54

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(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 54 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

ACGTTCCACA GCGATGAGAA CCAGAGAGAA AATGGATACT GTAATGGAGA CGCA

54

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 42 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

CTGCAGTATT TTGGCCCACT CTGTTTCATA TTCATATGCT AC

42

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 42 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CAAGCGAATG TATATCTTGA AGTAGCATAT GAATATGAAA CA

42

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 48 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

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(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

CTGCTCTGCC ACCTCACGGC CATGATCTCC ACCTGCGTCA ACCCCATC

45

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 48 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

GAAATTTTG TTCAGGAATC CATAAAAGAT GGGGTTGACG CAGGTGGA

48

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 47 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

TCATCGTCAC TTCTTACAGC ATTGAGACTG TCGTGGGGT CCTGGGT

47

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 46 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

ACAGTCACAC ACATCAGGCA GAGGTTACCC AGGACCCCCA CGACAG

46

(2) INFORMATION FOR SEQ ID NO:15:

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(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

TGCTTATCGC CAACCTGGCC TTCTCTGACT TCCTCATGTG CCTCC

45

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

TAGACGGCGG TCAGCGGCTG GCAGAGGAGG CACATGAGGA AGTCA

45

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

TGTCGGCCTT CATCCAGTGC ATGTCGGTGA CGGTCTCCAT CCTCT

45

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

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(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

CTCTCCAGGG CCACGAGGAC GAGCGAGAGG ATGGAGACCG TCACC

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

GCCTACCTGG GGATTGTGCT CATCTGGTC ATTGCCTGTG TCCTC

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

TGCTGTTGGC CAGGAAGGGC AGGGAGAGGA CACAGGCAAT GACCC

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

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CATCTACACC ACCTTCCTGC TCCTCTTCCA GTACTGCCTC CCACT

45

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

TGCATAACAG ACCAGGATGA AGCCCAGTGG GAGGCAGTAC TGGAA

45

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 46 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

CTGGTGGTGA TGGTGGTGGC CTTTGCCGTG CTCTGGCTGC CTCTGC

46

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

CAGTCTTCCA GGCTGTTGAA CACATGCAGA GGCAGCCAGA GCACG

45

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid

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(C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

ATCTTCTTAG TGTGCCACTT GCTTGCCATG GCCTCCACCT GCGTC

43

(2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 45 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

TGAGAAAGCC ATAGATGAAT GGGTTGACGC AGGTGGAGGC CATGG

45

(2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1439 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION: 178..1306

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

ATAGCTCTCA AGCCATAAGA TATAAGTAGC TAAGAATTGT CTCCCTCTCC CTGTCCTTG

50

TTCTTACCTG GTTCCATTTT ACATGCCTGG ACCTTTGAGT TCCATTGTT TGTTTTGCAG

120

GCTACACTCA GAAGTGGGCC CTTTAGTCTT GAAGTTCCTG GTCTTCTCAC ACCCACC

177

ATG AAT ACC TCT CAT CTC ATG GCC TCC CTT TCT CCG GCA TTC GAA CAA
 Met Asn Thr Ser His Leu Met Ala Ser Leu Ser Pro Ala Phe Leu Gln

225

1 5 10 15

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Gly Lys Asn Gly Thr Asn Pro Leu Asp Ser Leu Tyr Asn Leu Ser Asp	20	25	30	
GGC TGC CAG GAT TCG GCA GAT CTG TTG GCC TTC ATC ATC ACC ACC TAC				321
Gly Cys Gln Asp Ser Ala Asp Leu Leu Ala Phe Ile Ile Thr Thr Tyr	35	40	45	
AGC GTT GAG ACC GTC TTG GGG GTC CTA GGA AAC CTC TGC TTG ATA TTT				363
Ser Val Glu Thr Val Leu Gly Val Leu Gly Asn Leu Cys Leu Ile Phe	50	55	60	
GTG ACC ACA AGG CAA AAG GAA AAG TCC AAT GTG ACC AAC CTA CTC ATT				417
Val Thr Thr Arg Gln Lys Glu Lys Ser Asn Val Thr Asn Leu Leu Ile	65	70	75	
GCC AAC CTG GCC TTC TCT GAC TTC CTC ATG TGT CTC ATC TGC CAG CCG				463
Ala Asn Leu Ala Phe Ser Asp Phe Leu Met Cys Leu Ile Cys Gln Pro	85	90	95	
CTC ACG GTC ACC TAC ACC ATC ATG GAC TAC TGG ATC TTC GGC GAA GTC				513
Leu Thr Val Thr Tyr Thr Ile Met Asp Tyr Trp Ile Phe Gly Glu Val	100	105	110	
CTT TGC AAG ATG TTA ACG TTC ATC CAG TGT ATG TCG GTG ACA GTC TCC				561
Leu Cys Lys Met Leu Thr Phe Ile Gln Cys Met Ser Val Thr Val Ser	115	120	125	
ATC CTC TCA CTG GTC CTT GTG GCC CTG GAG AGG CAC CAG CTC ATT ATC				609
Ile Leu Ser Leu Val Leu Val Ala Leu Glu Arg His Gln Leu Ile Ile	130	135	140	
AAC CCG ACT GGC TGG AAA CCC AGC ATT TCC CAG GCC TAC CTG GGG ATT				657
Asn Pro Thr Gly Trp Lys Pro Ser Ile Ser Gln Ala Tyr Leu Gly Ile	145	150	155	
160				
GTG GTC ATC TGG TTC ATT TCT TGT TTC CTC TCC TTG CCC TTC CTG GCC				705
Val Val Ile Trp Phe Ile Ser Cys Phe Leu Ser Leu Pro Phe Leu Ala	165	170	175	
AAT AGC ATC CTG AAC GAC CTC TTC CAC TAC AAC CAC TCT AAG GTT GTG				753
Asn Ser Ile Leu Asn Asp Leu Phe His Tyr Asn His Ser Lys Val Val	180	185	190	
GAG TTT CTG GAA GAC AAG GTT GTC TGC TTT GTG TCC TGG TCC TCG GAT				801
Glu Phe Leu Glu Asp Lys Val Val Cys Phe Val Ser Trp Ser Ser Asp	195	200	205	
CAC CAC CGC CTC ATC TAC ACC ACC TTT CTG CTG CTC TTC CAA TAC TGC				849
His His Arg Leu Ile Tyr Thr Thr Phe Leu Leu Leu Phe Gln Tyr Cys	210	215	220	
GTC CCT CTG GCC TTC ATC CTG GTC TGC TAC ATG CGT ATC TAT CAG CGC				897
Val Pro Leu Ala Phe Ile Leu Val Cys Tyr Met Arg Ile Tyr Gln Arg	225	230	235	
240				
CTG CAG AGG CAG AGG CGT GCG TTC CAC ACG CAC ACT TGC AGC TCA CGA				945
Leu Gln Arg Gln Arg Arg Ala Phe His Thr His Thr Cys Ser Ser Arg	245	250	255	
GTG GGG CAG ATG AAG CGG ATC AAT GGC ATG CTC ATG GCA ATG GTG ACT				993
Val Glu Gln Met Lys Pro Ile Asn Gly Met Leu Met Ala Met Val Thr	260	265	270	
GCC TTT GCA GTT CTC TGG CTG CCC CTG CAT GTG TTC AAC ACT CTG GAG				1041
Ala Phe Ala Val Leu Trp Leu Pro Leu His Val Phe Asn Thr Leu Glu	275	280	285	
GAC TGG TAC CAG GAA GCC ATC CCT GCT TGC CAT GGC AAC CTC ATC TTC				1089
Asp Trp Tyr Gln Glu Ala Ile Pro Ala Cys His Gly Asn Leu Ile Phe	290	295	300	

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TTG ATG TGC CAC CTG TTT GGC ATG GCT TCC ACC TGT GTC AAC CCT TTC	1187
Leu Met Cys His Leu Phe Ala Met Ala Ser Thr Cys Val Asn Pro Phe	
305 310 315 320	
ATC TAT GGC TTT CTC AAC ATC AAC TTC AAG AAG GAC ATC AAG GCT CTG	1188
Ile Tyr Gly Phe Leu Asn Ile Asn Phe Lys Lys Asp Ile Lys Ala Leu	
325 330 335	
GTT CTG ACC TGC CGT TGC AGG CCA CCT CAA GGG GAG CCT GAG CCT CTG	1283
Val Leu Thr Cys Arg Cys Arg Pro Pro Gln Gly Glu Pro Glu Pro Leu	
340 345 350	
CCC CTG TCC ACT GTG CAC ACG GAC CTC TCC AAG GGA TCT ATG AGG ATG	1281
Pro Leu Ser Thr Val His Thr Asp Leu Ser Lys Gly Ser Met Arg Met	
355 360 365	
GGT AGC AAG TCT AAC GTC ATG TAG T CATGTCTAGG CTCTTCCGCC	1326
Gly Ser Lys Ser Asn Val Met	
370 375	
ATTTCTTCG ACACACCCCTT TCACTGAGCT AAGTAGACAC AATGCAAGCT GTGGTATCAT	1386
CCTGCCATTT CTGGTCTTG GGGCCCAGAC AGGCGGCAAG AGACTTGAAG CTT	1439

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 376 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Met Asn Thr Ser His Leu Met Ala Ser Leu Ser Pro Ala Phe Leu Gln	
1 5 10 15	
Gly Lys Asn Gly Thr Asn Pro Leu Asp Ser Leu Tyr Asn Leu Ser Asp	
20 25 30	
Gly Cys Gln Asp Ser Ala Asp Leu Leu Ala Phe Ile Ile Thr Thr Tyr	
35 40 45	
Ser Val Glu Thr Val Leu Gly Val Leu Gly Asn Leu Cys Leu Ile Phe	
50 55 60	
Val Thr Thr Arg Gln Lys Glu Lys Ser Asn Val Thr Asn Leu Leu Ile	
65 70 75 80	
Ala Asn Leu Ala Phe Ser Asp Phe Leu Met Cys Leu Ile Cys Gln Pro	
85 90 95	
Leu Thr Val Thr Tyr Thr Ile Met Asp Tyr Trp Ile Phe Gly Glu Val	
100 105 110	
Leu Cys Lys Met Leu Thr Phe Ile Gln Cys Met Ser Val Thr Val Ser	
115 120 125	
Ile Leu Ser Leu Val Leu Val Ala Leu Glu Arg His Gln Leu Ile Ile	
130 135 140	
Asn Pro Thr Gly Trp Lys Pro Ser Ile Ser Gln Ala Tyr Leu Gly Ile	
145 150 155 160	
Val Val Ile Trp Phe Ile Ser Cys Phe Leu Ser Leu Pro Phe Leu Ala	
165 170 175	

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180	185	190
Glu Phe Leu Glu Asp Lys Val Val Cys Phe Val Ser Trp Ser Ser Asp		
195	200	205
His His Arg Leu Ile Tyr Thr Thr Phe Leu Leu Phe Gln Tyr Cys		
210	215	220
Val Pro Leu Ala Phe Ile Leu Val Cys Tyr Met Arg Ile Tyr Gln Arg		
225	230	235
Leu Gln Arg Gln Arg Arg Ala Phe His Thr His Thr Cys Ser Ser Arg		
245	250	255
Val Gly Gln Met Lys Pro Ile Asn Gly Met Leu Met Ala Met Val Thr		
260	265	270
Ala Phe Ala Val Leu Trp Leu Pro Leu His Val Phe Asn Thr Leu Glu		
275	280	285
Asp Trp Tyr Gln Glu Ala Ile Pro Ala Cys His Gly Asn Leu Ile Phe		
290	295	300
Leu Met Cys His Leu Phe Ala Met Ala Ser Thr Cys Val Asn Pro Phe		
305	310	315
Ile Tyr Gly Phe Leu Asn Ile Asn Phe Lys Lys Asp Ile Lys Ala Leu		
325	330	335
Val Leu Thr Cys Arg Cys Arg Pro Pro Gln Gly Glu Pro Glu Pro Leu		
340	345	350
Pro Leu Ser Thr Val His Thr Asp Leu Ser Lys Gly Ser Met Arg Met		
355	360	365
Gly Ser Lys Ser Asn Val Met *		
370	375	